

# Effects of Ocean Acidification on Marine Primary Producers

**Kunshan Gao** et al.

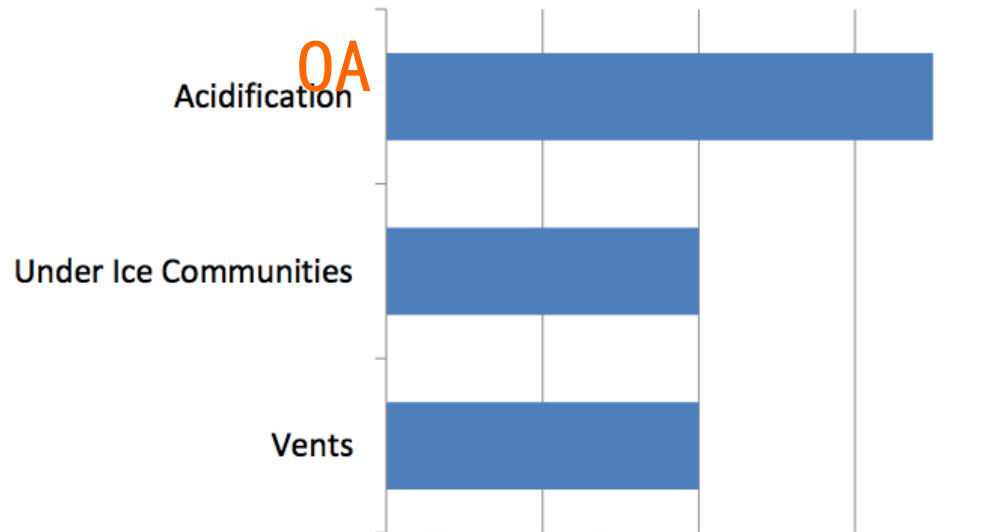
PICES, Qingdao, 2015



**State Key Laboratory of Marine Environmental Science (Xiamen Univ.)**

Xiamen University

**What are the three most important phenomena to explore?**



Google Search (<1 s)

2009年 “海洋酸化” /ocean acidification: 70/700 K

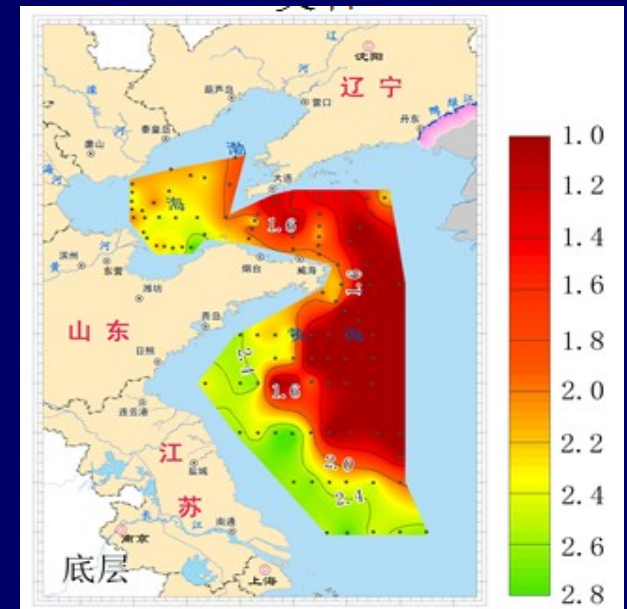
2013年 “海洋酸化” /ocean acidification: 2.58/2.59 M

“Chinese OA awareness index”

# Documented low pH in the Chinese coastal waters

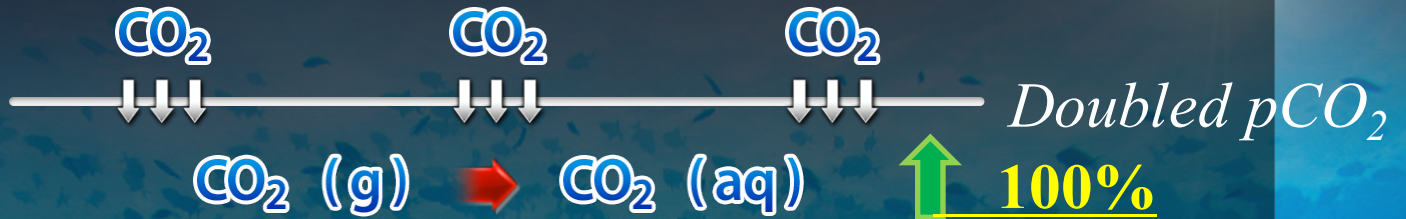
Regions	Low pH
The Bohai Sea ( <i>Chinese Science Bulletin</i> 2012)	7.64 H <sup>+</sup> rise by 220%
The Yellow Sea ( <i>Biogeosciences</i> 2014)	7.80
The East China Sea ( <i>Biogeosciences</i> 2013)	7.80
The Northern South China Sea ( <i>JGR</i> 2011)	7.9

Ocean acidification is occurring in the Chinese waters.



“中国海洋环境公报 2012”

# Seawater Carbonate System Changes



0.3-0.4,                      40%,                      10%

\text{CO}\_3^{2-} ↓       $\text{HCO}_3^-$  ↑

$\Omega = \frac{[\text{CO}_3^{2-}]_{\text{MEAS}}}{[\text{CO}_3^{2-}]_{\text{CAL}}} \downarrow \text{40\%}$

- Briefing Ecological Effects of OA
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G Caramna

**SEA  
CHANGE  
WITH  
PLYMOUTH  
UNIVERSITY**



**UK Ocean Acidification  
Research Programme**



**save our seas  
FOUNDATION**

# Effects on Ecosystems

## A window into the future of coral reefs?

pH 8.05: Today

pH 7.95: ~ year 2050

pH 7.8: ~ year 2100



Fabricius et al. (2010) *Nat Clim Change*

Inoue et al. 2013 *Nat Clim Change*



Munday et al. 2009

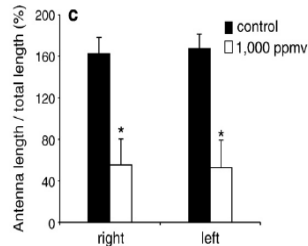
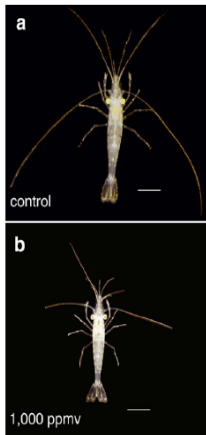
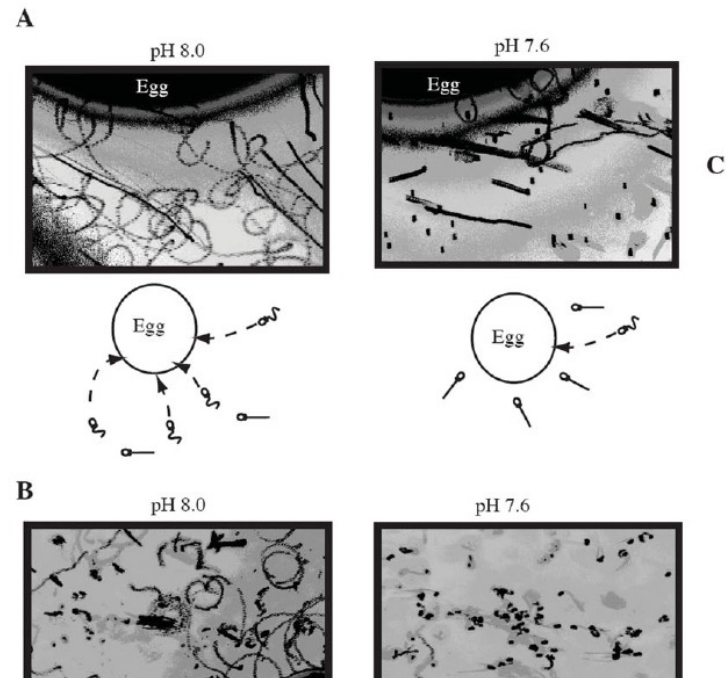
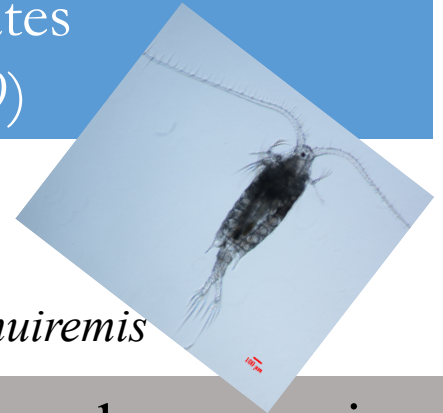


Fig. 6. Effects of 30-week exposure to 1,000 ppmv CO<sub>2</sub> on the antenna length of the marine shrimp *Palaeomon pacificus* (a control, b experimental). The ratio of antenna/total length was significantly smaller in experimental shrimps (54.0±21.9%, n=9, 2 individuals not measured) than in controls (164.9±14.2%, n=18, t-test, p<0.001, c). Bar=1 cm.



Reduce the sperm flagella motility of reef invertebrates (Morita et al.2009)



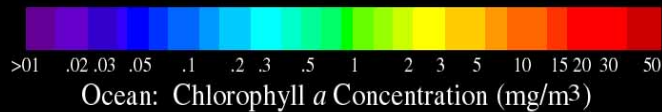
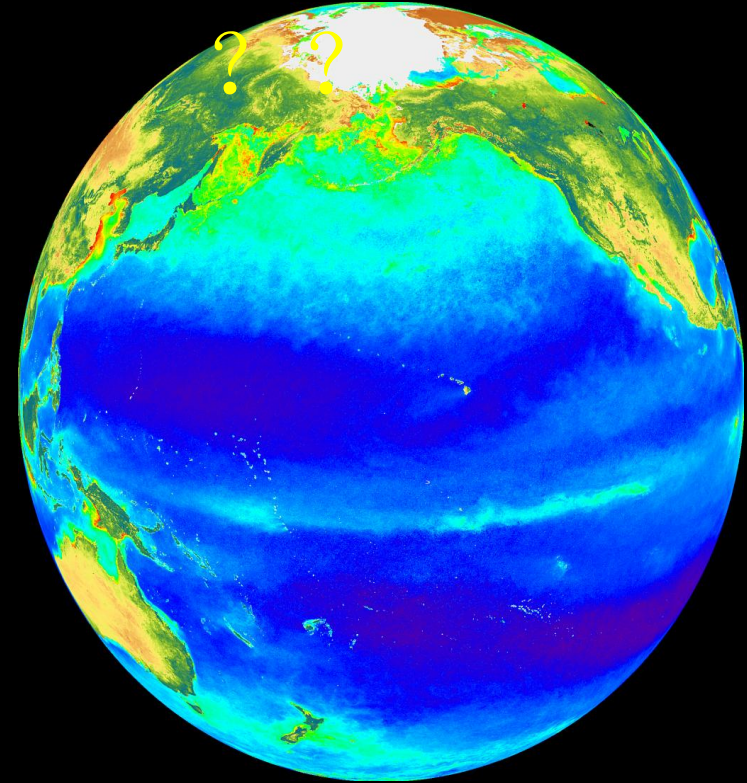
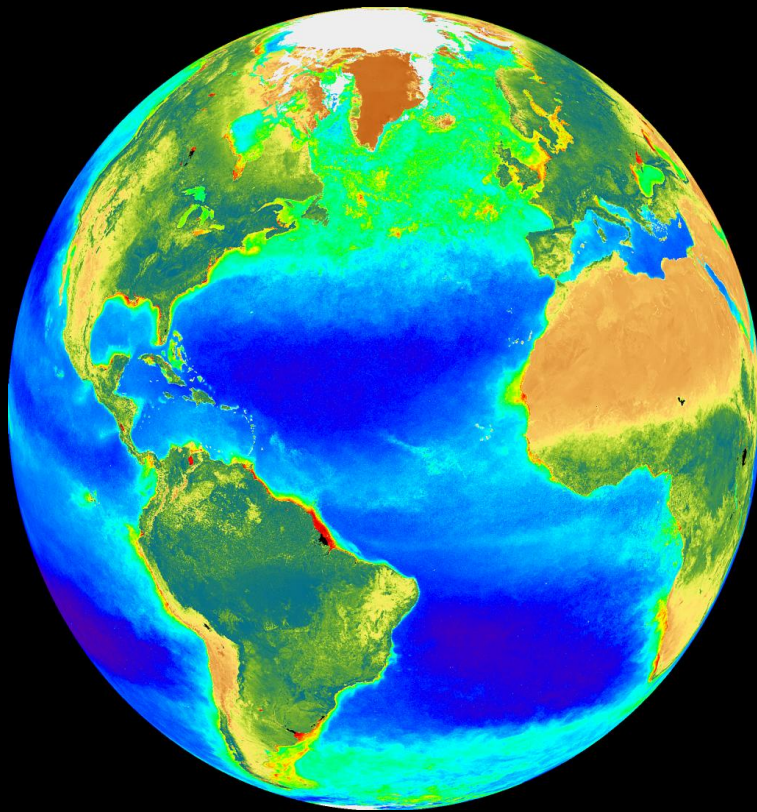
*Centropages tenuiremis*

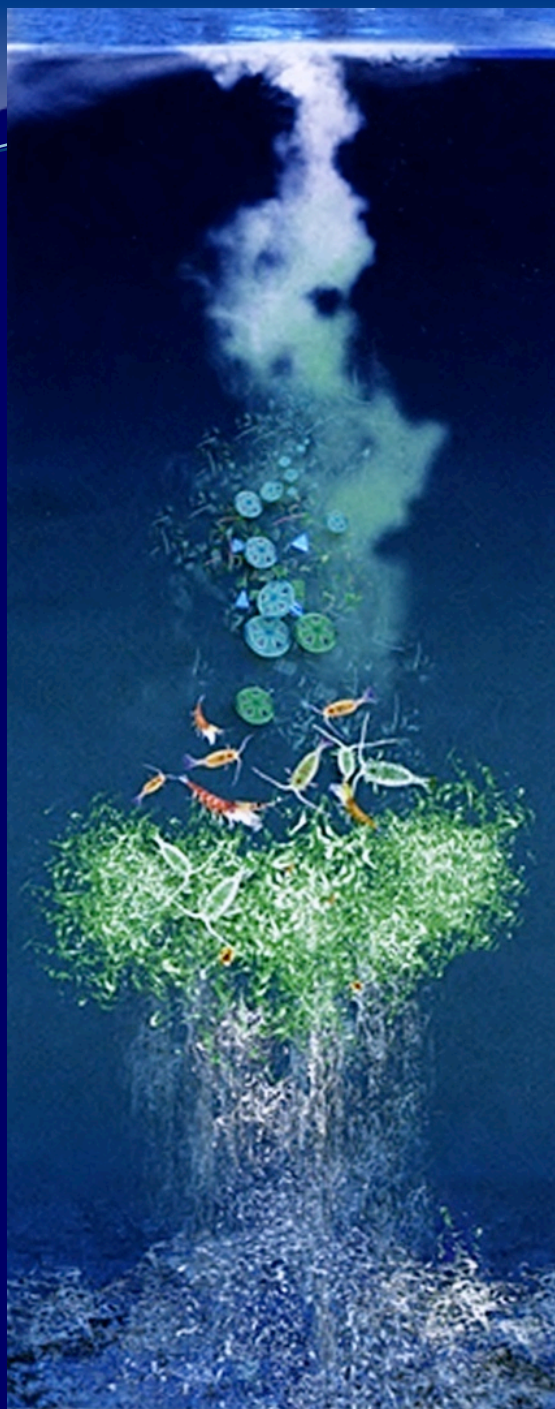
A marine secondary producer respirees and feeds more in a high CO<sub>2</sub> ocean  
Li & Gao 2012

Shorter antennae  
Kurihara et al.2008



Phytoplankton communities will change due to climatically-driven ocean changes (Falkowski et al.2007).  
Are marine PP going to sustain by **50%**





Photosynthesis-driven  
biological CO<sub>2</sub> pump  
would lead to more or  
less CO<sub>2</sub> dissolution into  
the oceans ?



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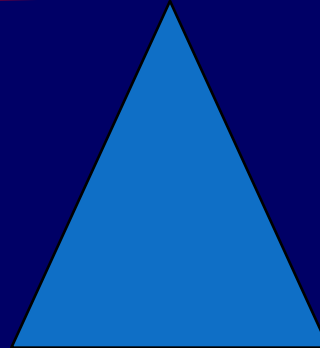
About 451 papers on phytoplankton and 282 macro algae's response to OA till Jul. 13, 2015 (OA-ICC bibliographic database)

*Nature 1997,*  
*Science 2008.....*

*Nature 2000, 2011;*  
*Nat Clim Change 2012*  
.....

• Stimulating                      Neutral                      Inhibitive

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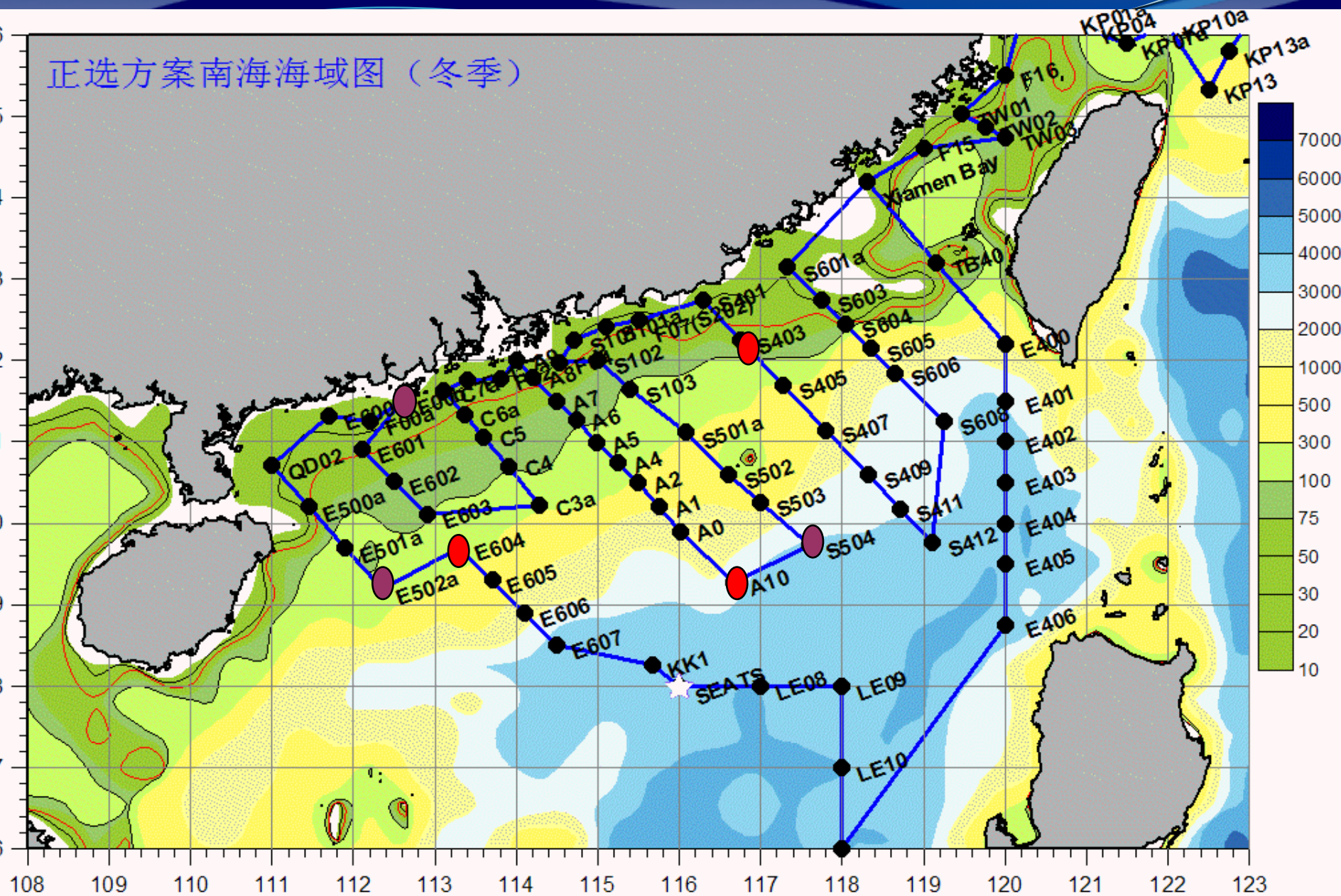
Growth

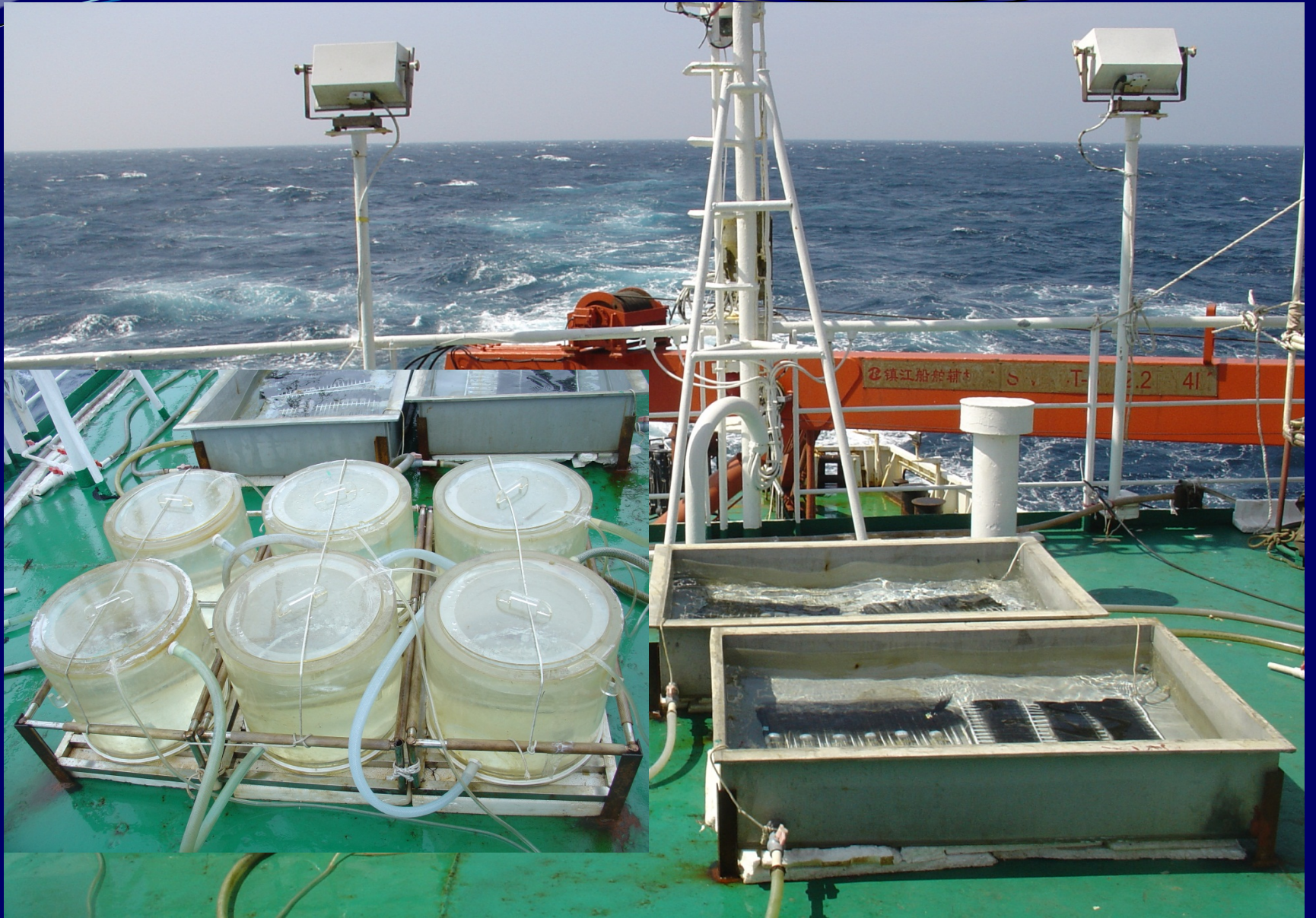
Photosynthesis

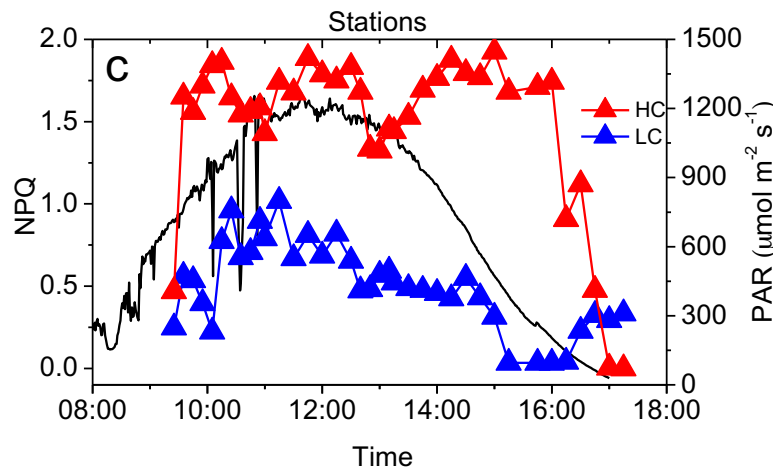
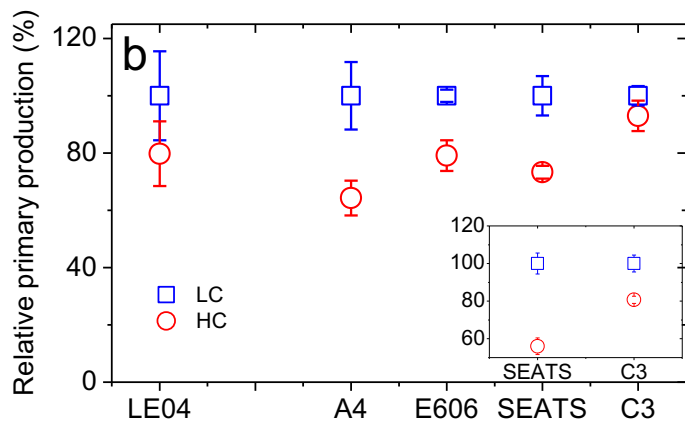
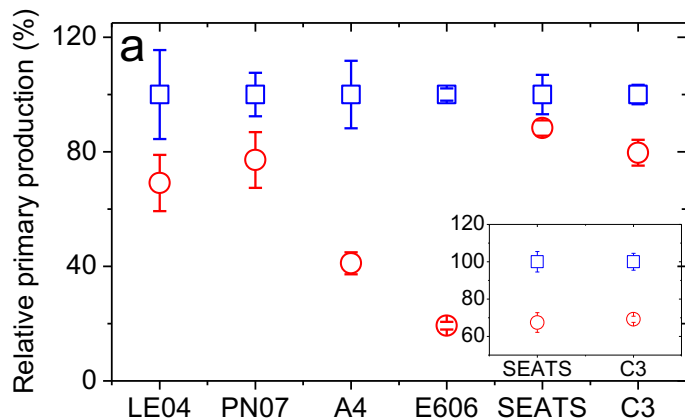
Respiration

Calcification

正选方案南海海域图（冬季）







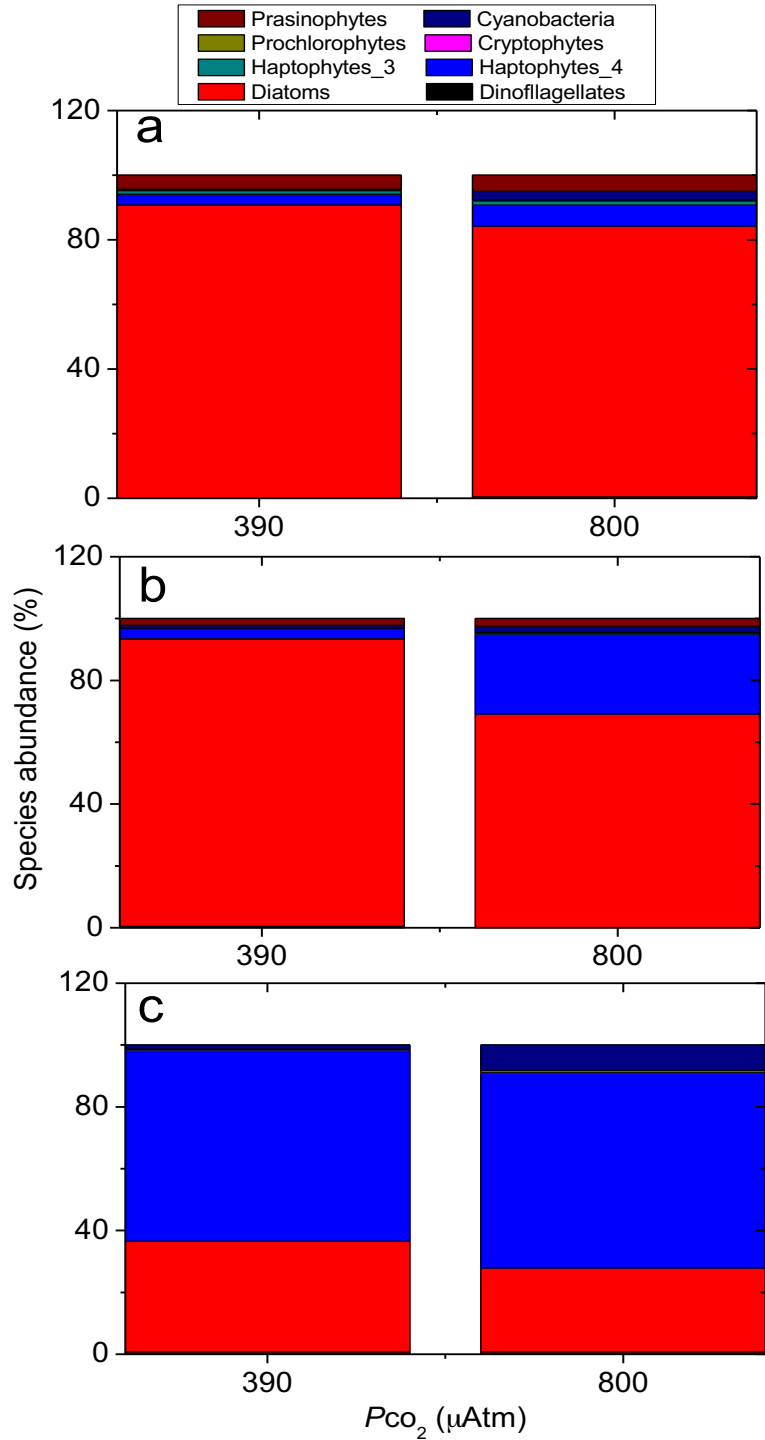
Growth at the high pCO<sub>2</sub> level thus significantly (P values are given in Supplementary Table 3) reduced primary productivity by 12-81% per volume of seawater (Fig. 1a) and by 7-36% per amount of chl. *a* (Fig. 1b) based on the daytime 6 or 12 h <sup>14</sup>C-traced incubations.

**Primary production** in CO<sub>2</sub> perturbed microcosms by phytoplankton assemblages collected in the South China Sea and East China Sea (station PN07). a) per volume of seawater ( $\mu\text{g C L}^{-1}\text{h}^{-1}$ ); b) per chlorophyll *a* ( $\mu\text{g C chl a}^{-1}\text{h}^{-1}$ ); Chl *a* concentration at PN07 was not measured. For the **high pCO<sub>2</sub> (HC, 800  $\mu\text{atm}$ )** for all stations except SEATS and C3, where 1000  $\mu\text{atm}$  pCO<sub>2</sub> was applied) and **low CO<sub>2</sub> (LC, 385  $\mu\text{atm}$ )** experiments, triplicate microcosms (32 L) were used for each pCO<sub>2</sub> level (seawater carbonate system parameters are given in Supplementary Table 1). The phytoplankton assemblages in all microcosms were equally exposed to 91% incident solar visible radiation. Detailed information for the stations and related physicochemical and biological features are given in Supplementary Table 2. Inset: additional 24 h-incubations carried out at the two stations. Error bars represent standard deviations of triplicate incubations of samples from triplicate microcosms. Details about the statistical comparisons among the treatments are given in Supplementary Table 3. Note, higher photosynthetic carbon fixation rates were found under low CO<sub>2</sub>

**Gao et al. 2012**

**Nature Climate Change**

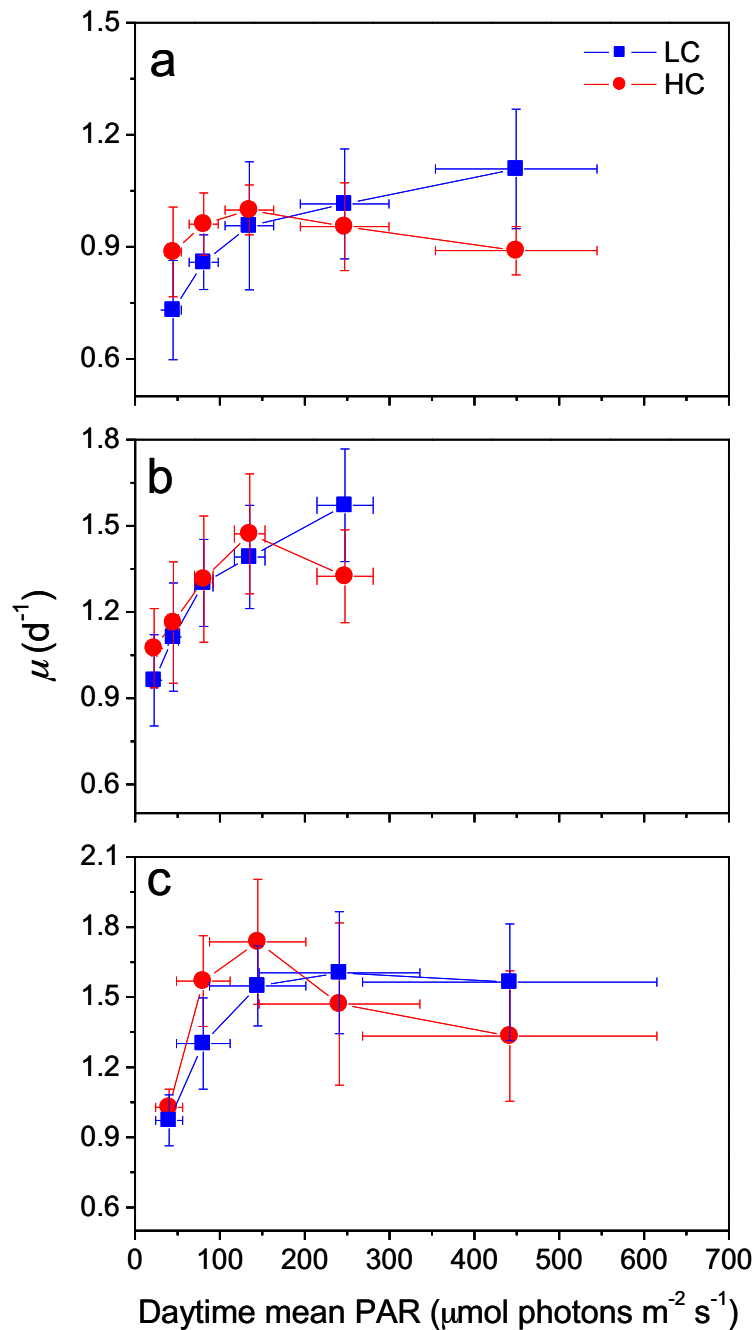
...anching (NPQ) ... E606 grown ... (gle) and high ... cosms on day ... 6. NPQ in other stations showed similar patterns (Supplementary Fig.2). The black line represents the visible light intensity of that day. Note, higher NPOs



Composition of phytoplankton assemblages grown under LC ( $pCO_2$  385  $\mu\text{atm}$ ) and HC ( $pCO_2$  800  $\mu\text{atm}$ ) levels. **a**. Station A4, January 2010, 7-day  $CO_2$  pre-conditioning; **b**. Station of A4, November 2010, 7-day  $CO_2$  pre-conditioning; **c**. Station E606, November 2010, 6-day  $CO_2$  pre-conditioning.

*Gao et al. 2012 Nature Climate Change*

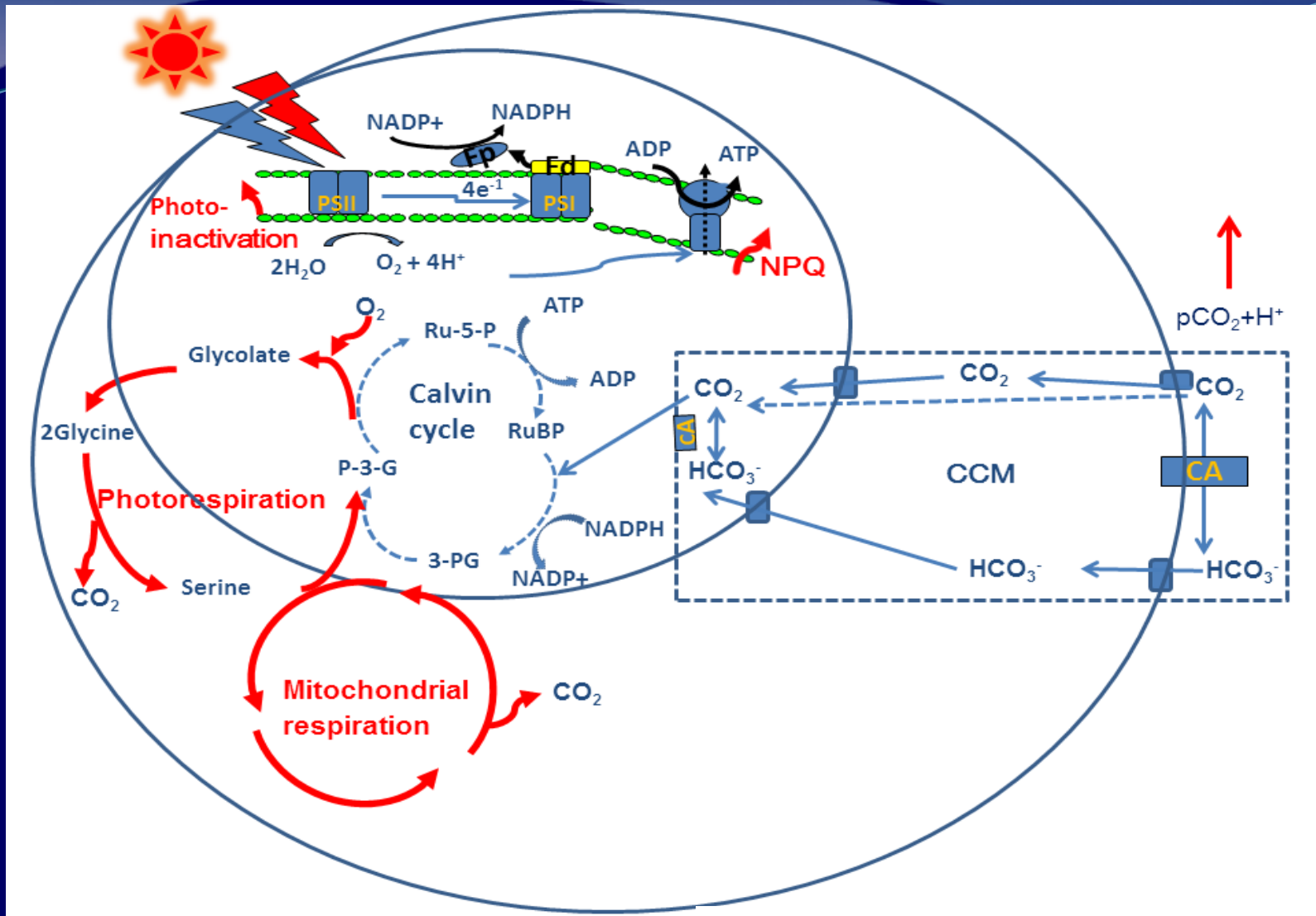




**Growth rate reversed at higher PAR levels, with the PAR thresholds (daytime mean PAR levels) at the reversion points being about 160, 125 and 178  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  for *P. tricornutum*, *T. pseudonana* and *S. costatum*, respectively.**

**These light levels correspond to 22-36 % of incident surface solar PAR levels and are equivalent to PAR levels at 26-39 m depth in the South China Sea**

Red: up-regulated

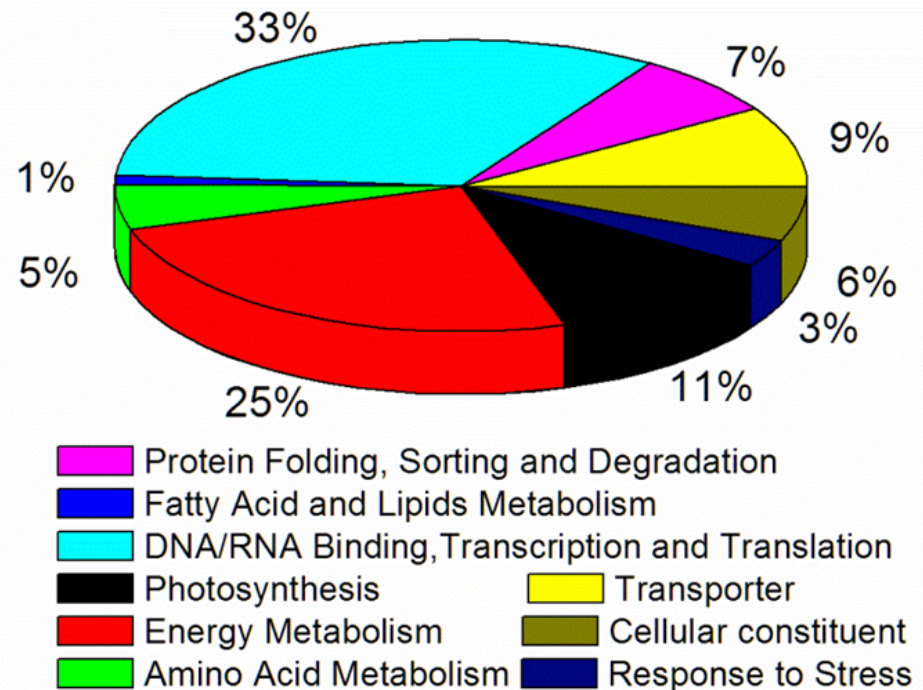


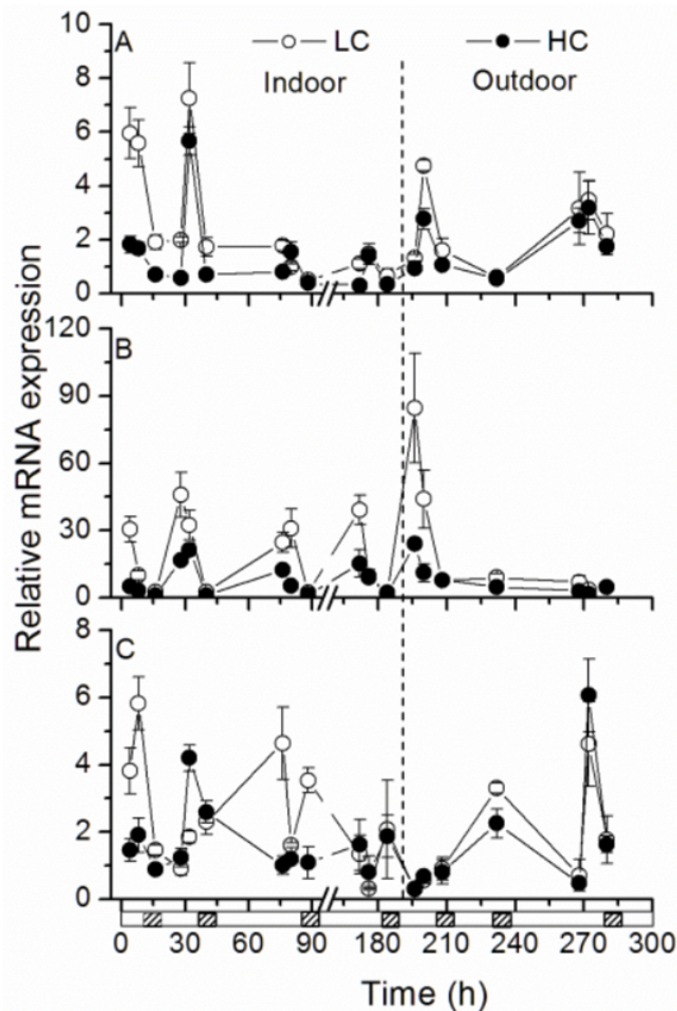
Gao et al. 2012  
Gao & Campbell 2014

**Down-regulated CCM**  
saving by 20% energy demand  
Hopkinson *et al.* 2011

# Genetic responses to OA

*Phaeodactylum tricornutum*





FCP

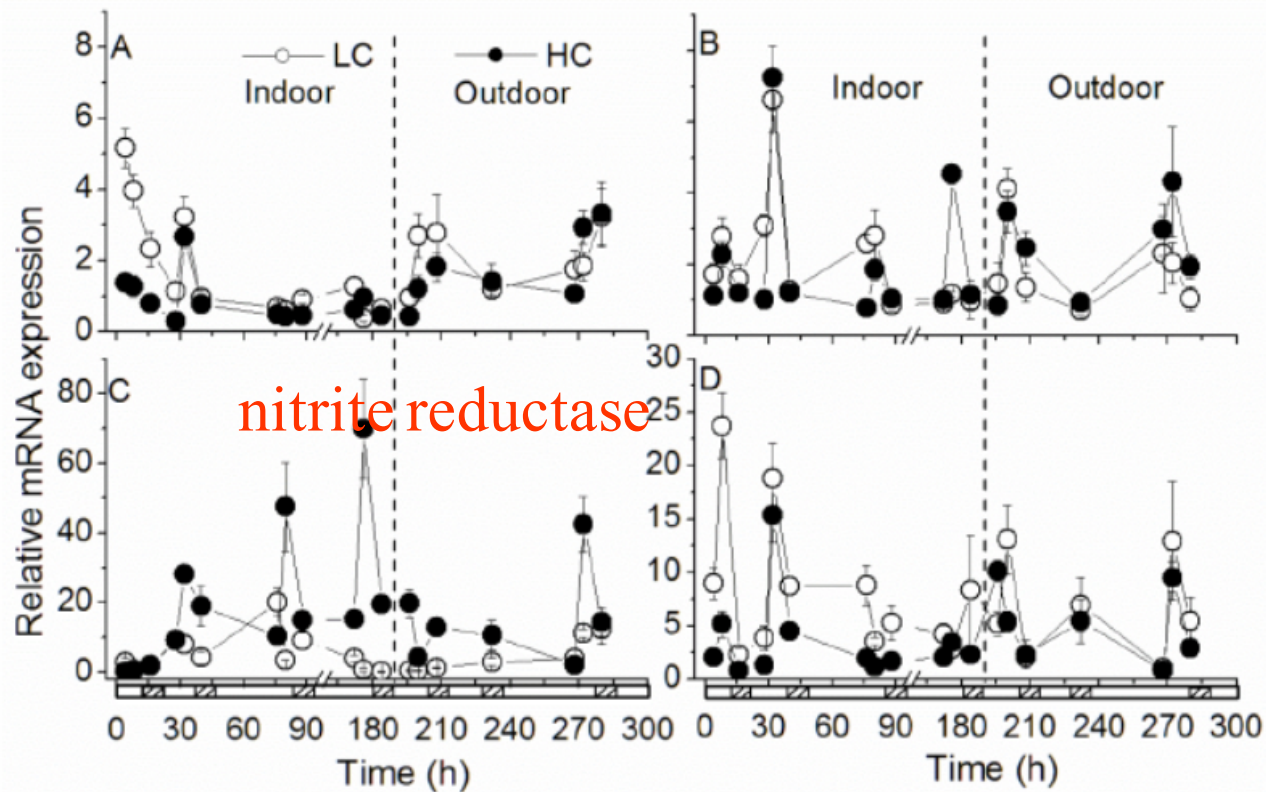
CA

RbcL

Fig.5 Time series of the relative abundances of fucoxanthin chlorophyll a/c protein, lhcf type (FCP; A), carbonic anhydrase (CA; B), ribulose-1, 5-bisphosphate carboxylase/oxygenase large subunit gene (RbcL; C) by quantitative real-time PCR (qPCR) of *P. tricornutum* cells grown in ambient (390  $\mu$ atm) and elevated CO<sub>2</sub> (1000  $\mu$ atm) conditions under constant light (indoor) and fluctuating sunlight levels (outdoor). Data are presented as means  $\pm$  SD, n = 3 (triplicate cultures).

*Li et al. 2015*

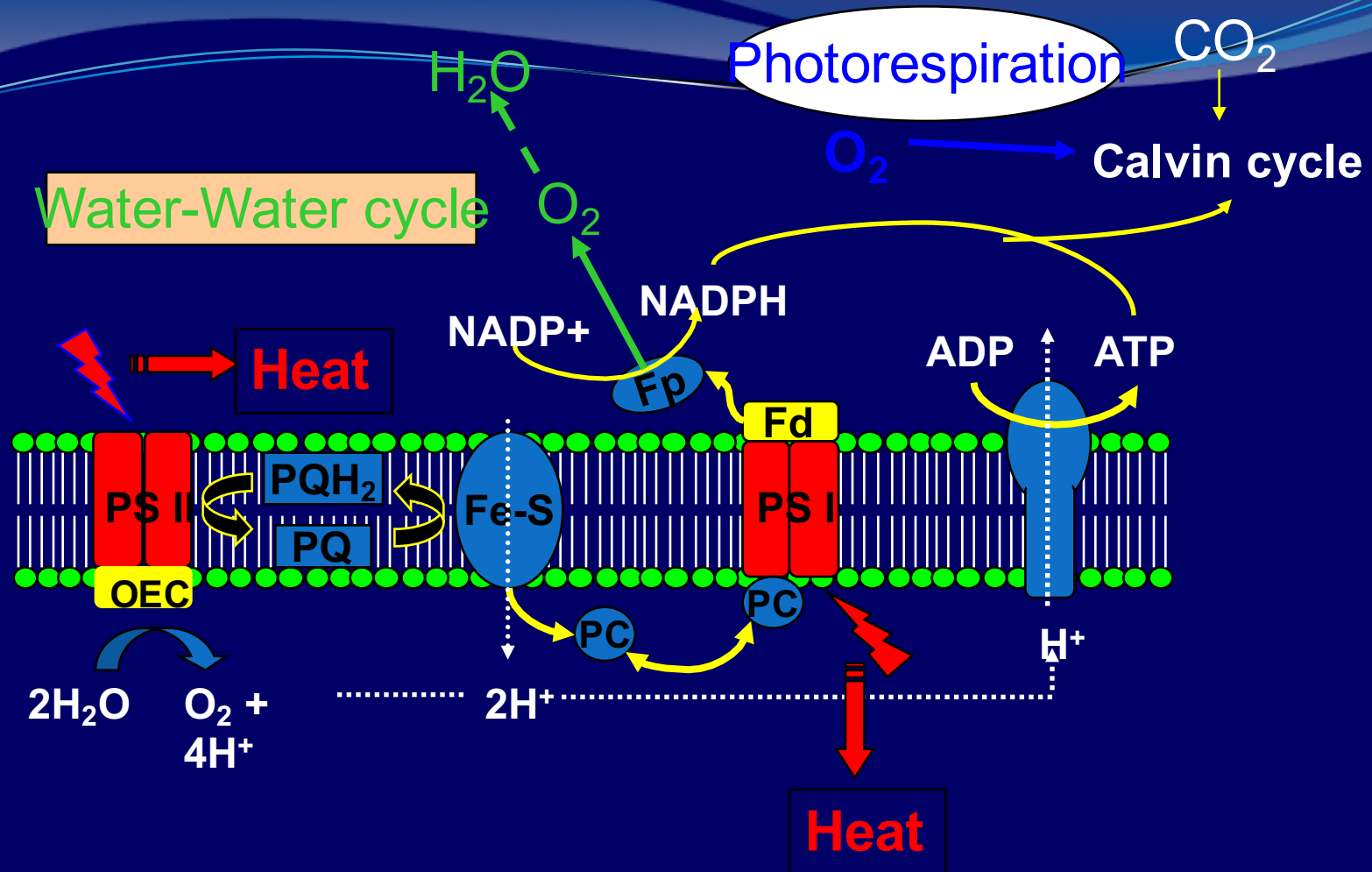
*Biogeosciences-discuss*



**Fig.6** The time series of the relative abundances of mitochondrial ATP synthase (mtATP; A), peroxisomal membrane protein-related (PMP; B), nitrite reductase (NiR; C) and NADH dehydrogenase subunit 2 (Ndh2; D) by quantitative real-time PCR (qPCR) of *P. tricornutum* cells grown in ambient (390 μatm) and elevated CO<sub>2</sub> (1000 μatm) conditions under constant light (indoor) and fluctuating sunlight levels (outdoor). Data are presented as means ± SD (triplicate cultures).

*Li et al. 2015*

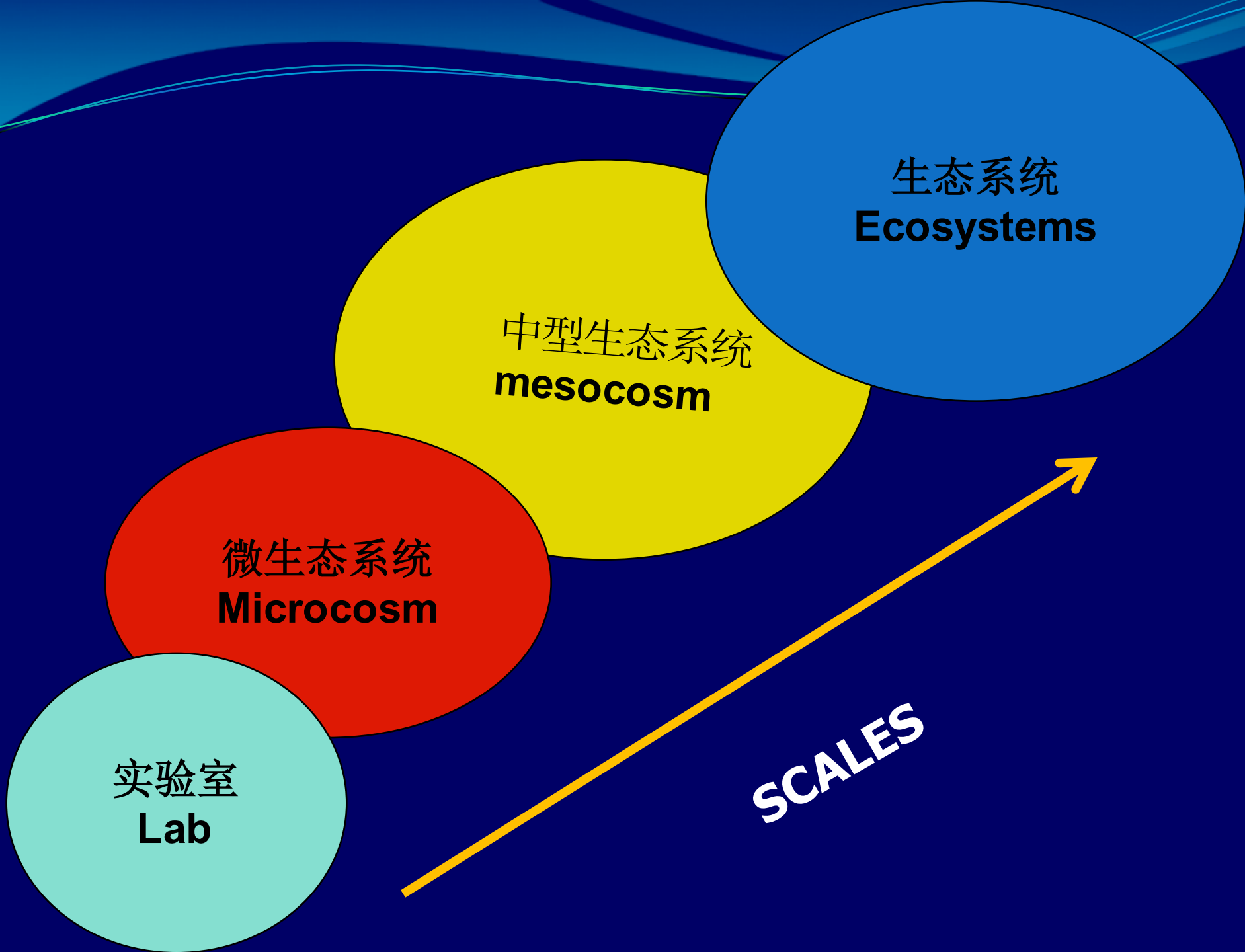
*Biogeosciences-discuss*



OA exacerbates high-light stress, stimulate photorespiration and mitochondrial respiration (diatoms, a green alga, a coccolithorpore)

*Wu et al. 2010 Biogeosciences;*  
*Gao et al. 2012a,b MEPS, Nat Clim Change;*  
*Xu and Gao 2013 Plant Physiol.*  
*Jin et al. 2015 Nat. Comm.*

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实验室  
Lab

微生态系统  
Microcosm

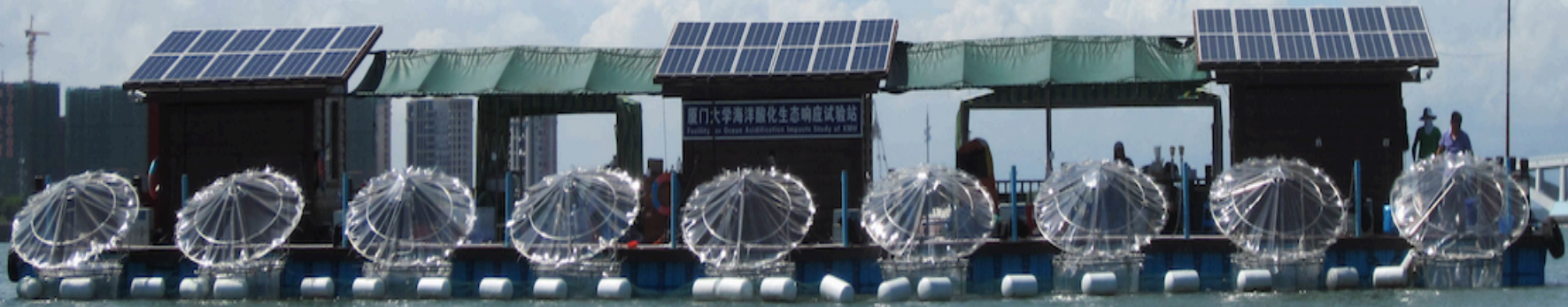
中型生态系统  
mesocosm

生态系统  
Ecosystems

SCALES

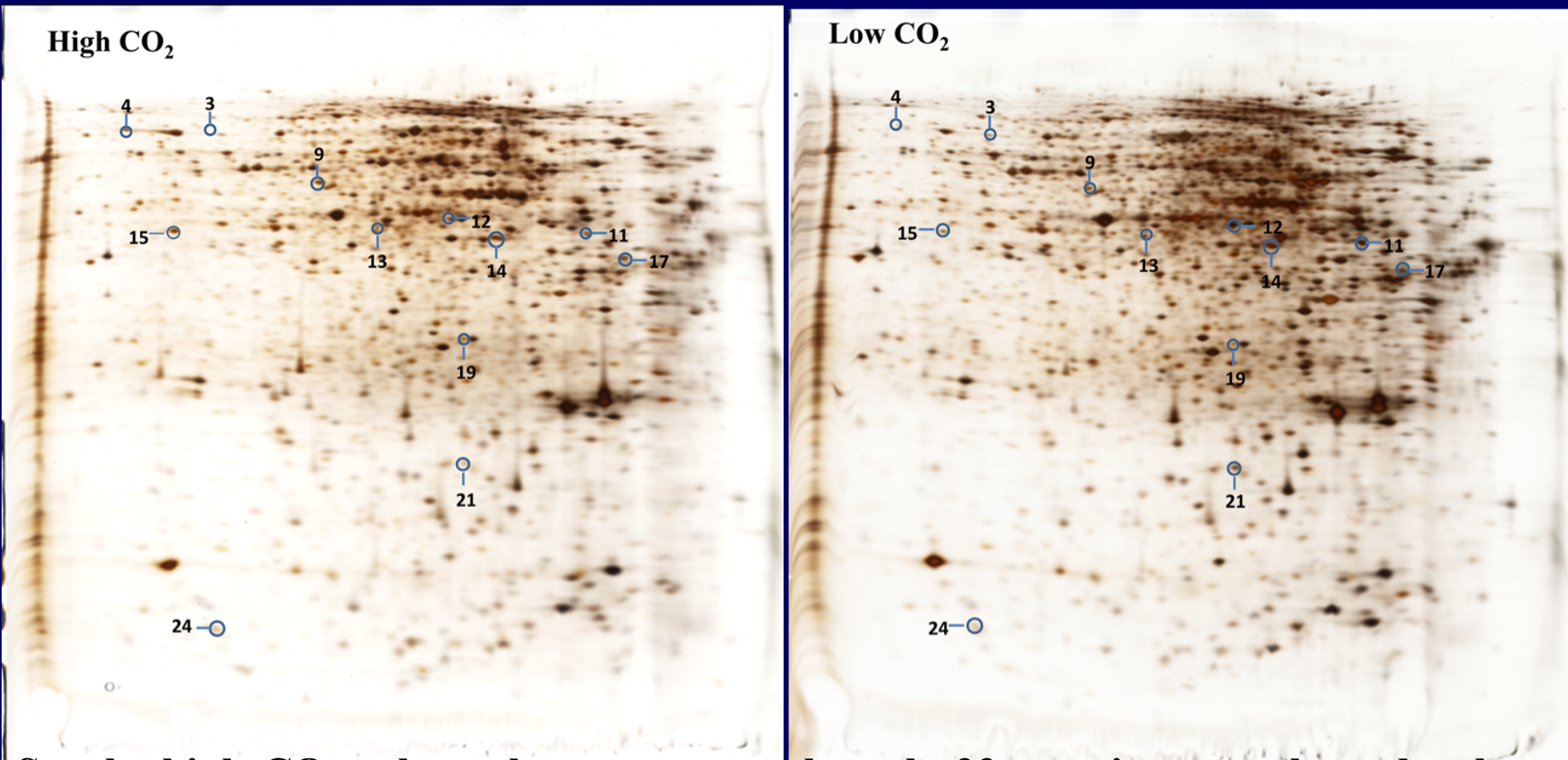


# Mesocosm Facility for Ocean Acidification Impact Study (FOANIC-XMU)



**FOANIC-XMU**  
**(N24° 31'48", E118° 10'47")**

# Proteomics responses, *Emiliana huxleyi* CCMP 1516

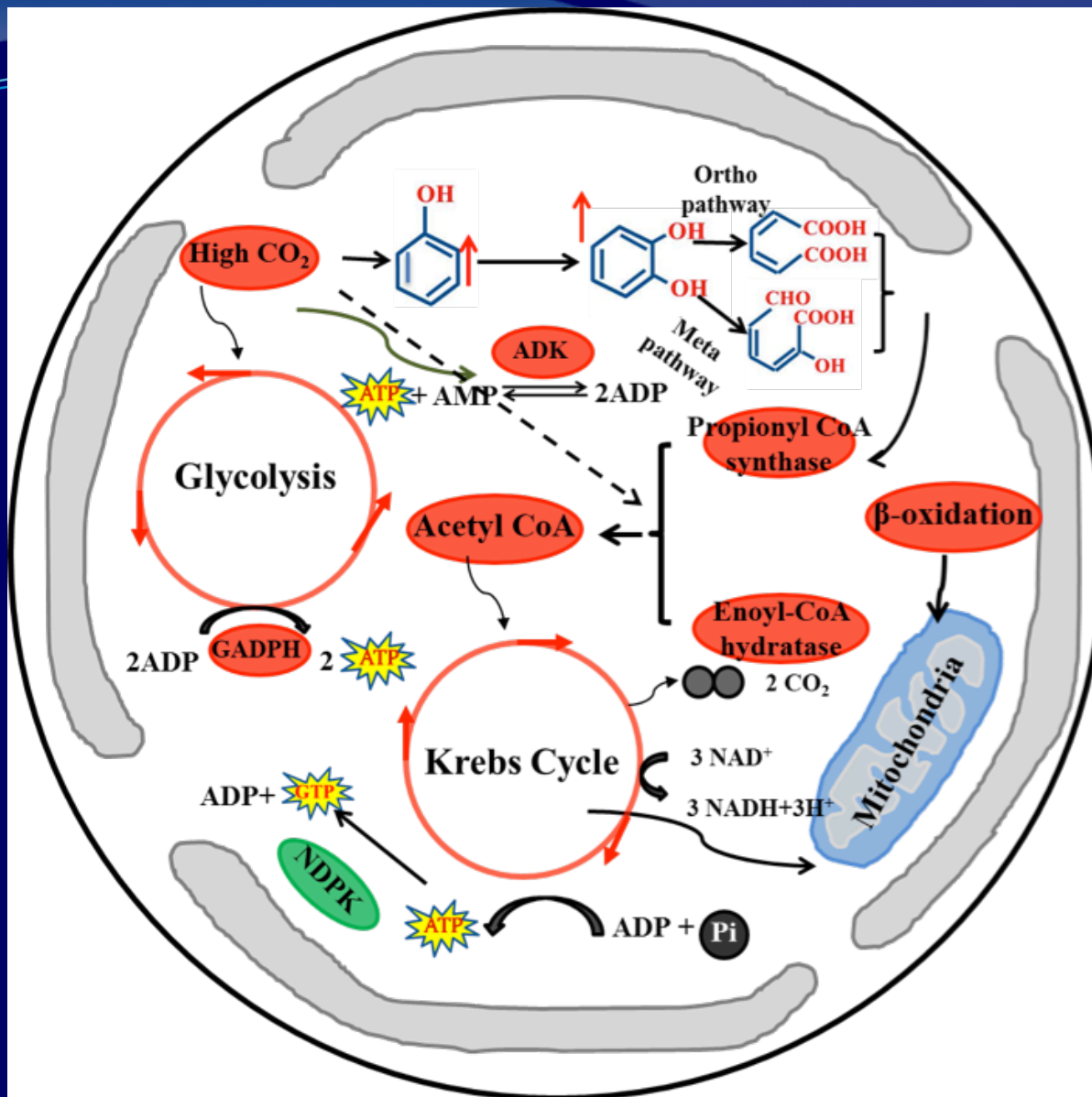


Set the high-CO<sub>2</sub> gels as the reference gels, only 33 protein spots showed at least 2-fold differences in expression level of the low-CO<sub>2</sub> gels. All the different protein spots are observed in the gels of different experiment conditions.

*Jin et al. 2015 Nat. Comm.*

# Various proteins, that showed statistically significant alterations in abundance greater than 2-fold, in HC and LC treatments

Spot Id	Protein identity	GI number	Protein score C.I. (%)	Total Ion C. I. %	Protein score (peptides)	MW/pI	Fold change		Function
							High CO <sub>2</sub>	Low CO <sub>2</sub>	
3	Propionyl CoA synthase	239994558	100	100	357(14)	69708.5/5.51	2.33	1.00	β-oxidation
4	Serine protein kinase	239995429	100	99.946	177(15)	74347.3/5.31	2.82	1.00	Protein kinase, signal transduction
9	Hypothetical protein AmacA_2	223994739	100	100	805(22)	51069.6/5.61	2.01	1.00	Unknown
11	Hypothetical protein MDMS009_211	254489880	100	100	440(11)	447891.1/4.87	1.00	4.34	Unknown
12	Methane/ phenol/ toluene hydroxylase	148260382	100	100	238(5)	39315.7/5.76	3.40	1.00	Phenol biodegradation
13	Acyl-CoA dehydrogenase family protein	83943662	100	100	438(18)	44108.4/5.55	1.00	2.66	β-oxidation
14	Chloroplast glyceraldehyde-3-phosphate dehydrogenase	77024139	100	100	336(7)	44096.1/5.2	2.92	1.00	Glycolysis
15	Conserved hypothetical protein (bacterium S5)	288797257	100	99.996	166(7)	21306.1/4.87	2.50	1.00	Unknown

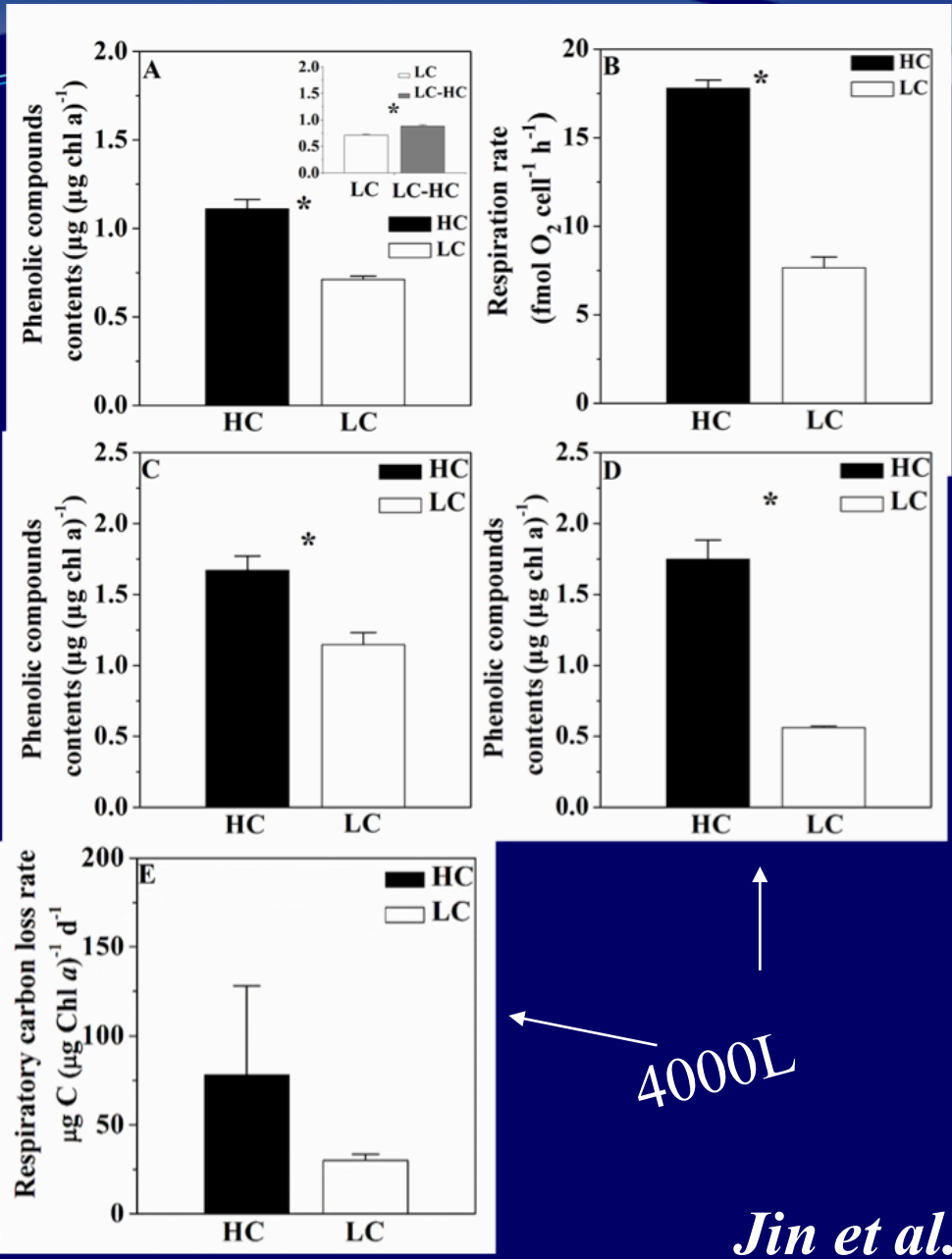


Red:  
up-regulated

Metabolic pathways in the coccolithophorid *Emiliana huxleyi* altered under ocean acidification (HC, CO<sub>2</sub> 1,000 matm; pH<sub>NBS</sub> 7.81) based on proteomic, physiological and biochemical analyses.

Lab

30L

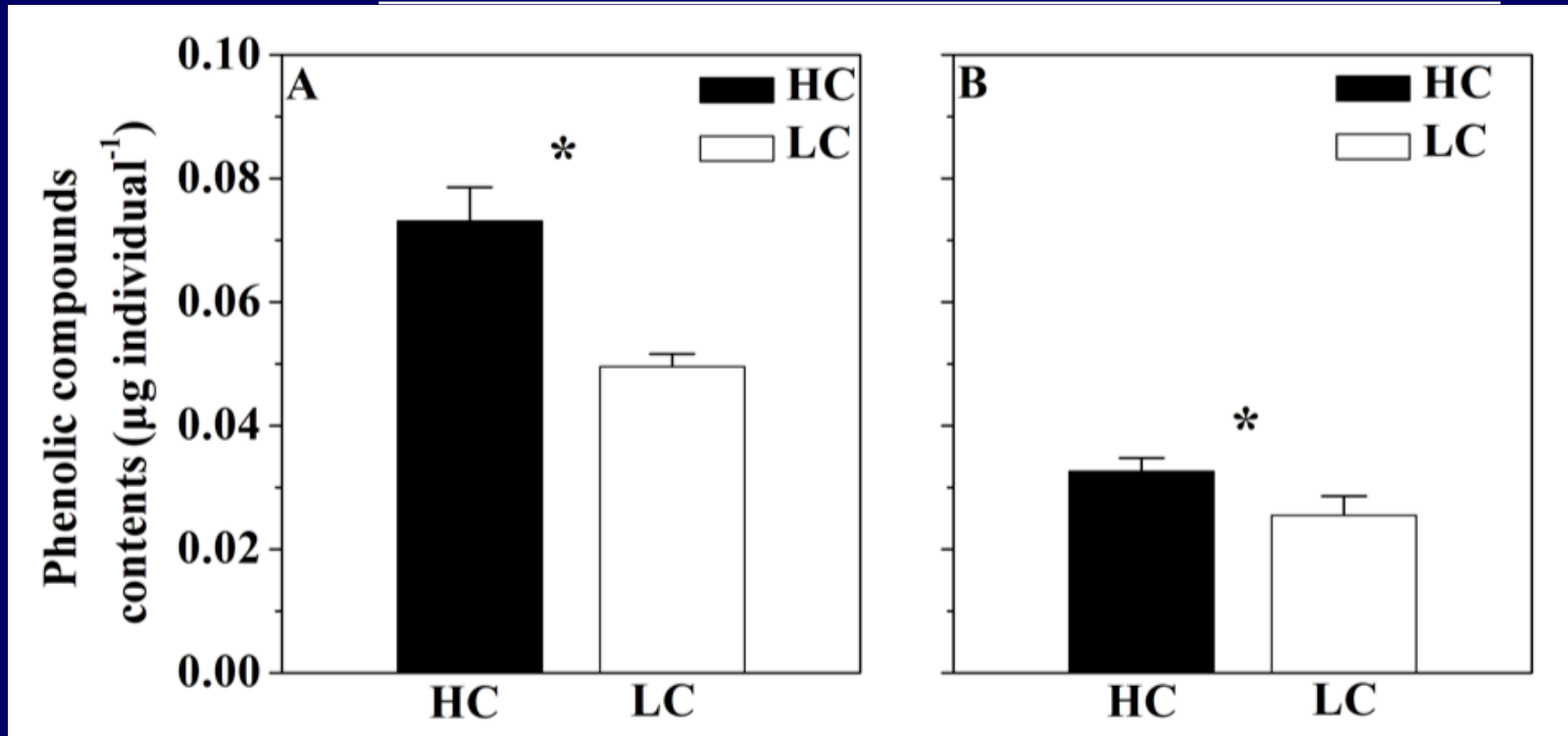


4000L

Jin et al. Nature Comm. 2015

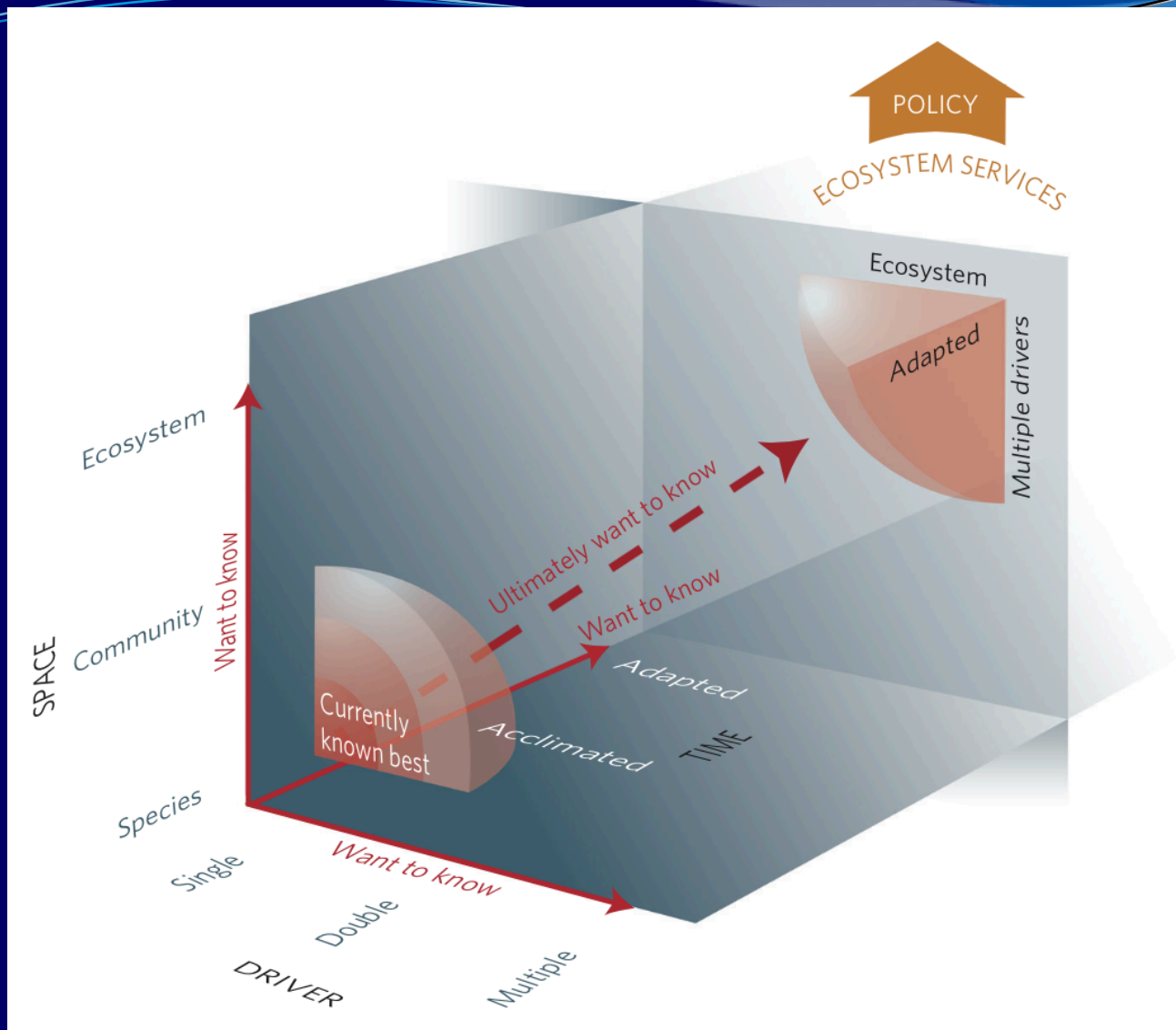
30 L

4000 L



Content of phenolic compounds (mg per individual) in zooplankton assemblages (body size >112  $\mu\text{m}$ )

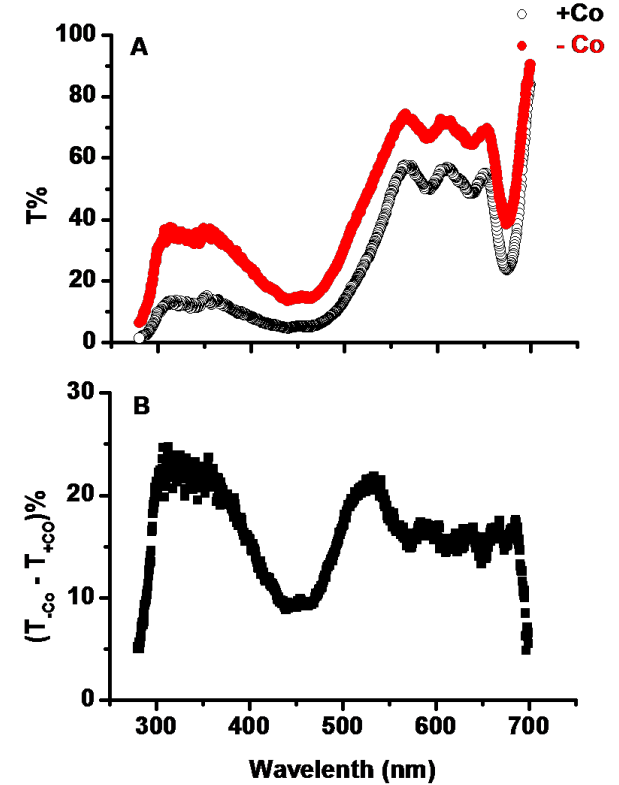
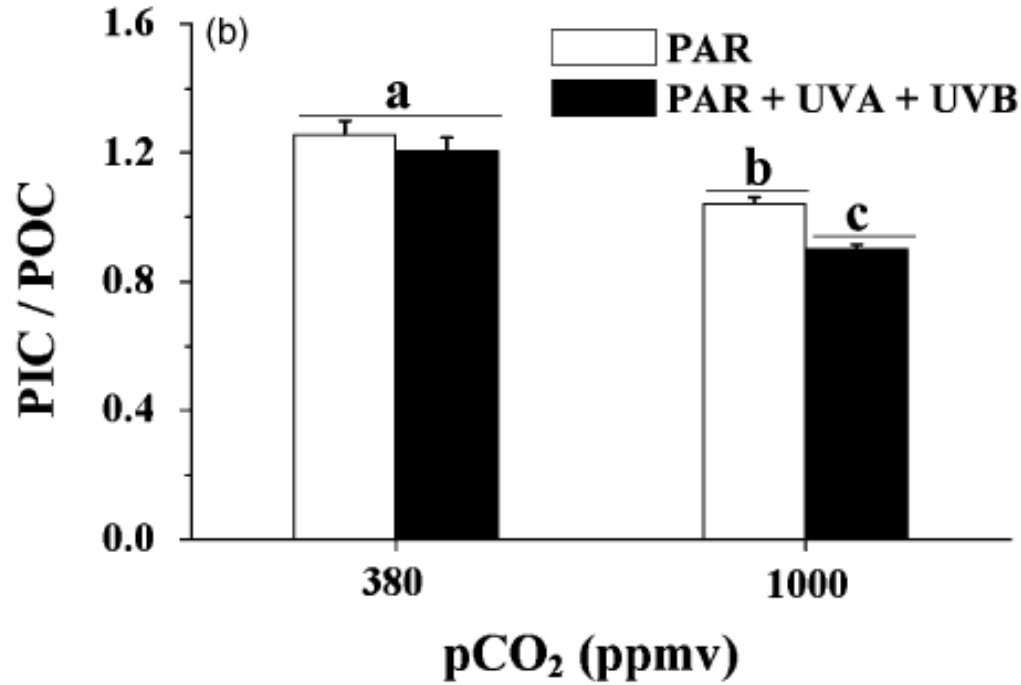
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*Riebesell and Gattuso (2015)*

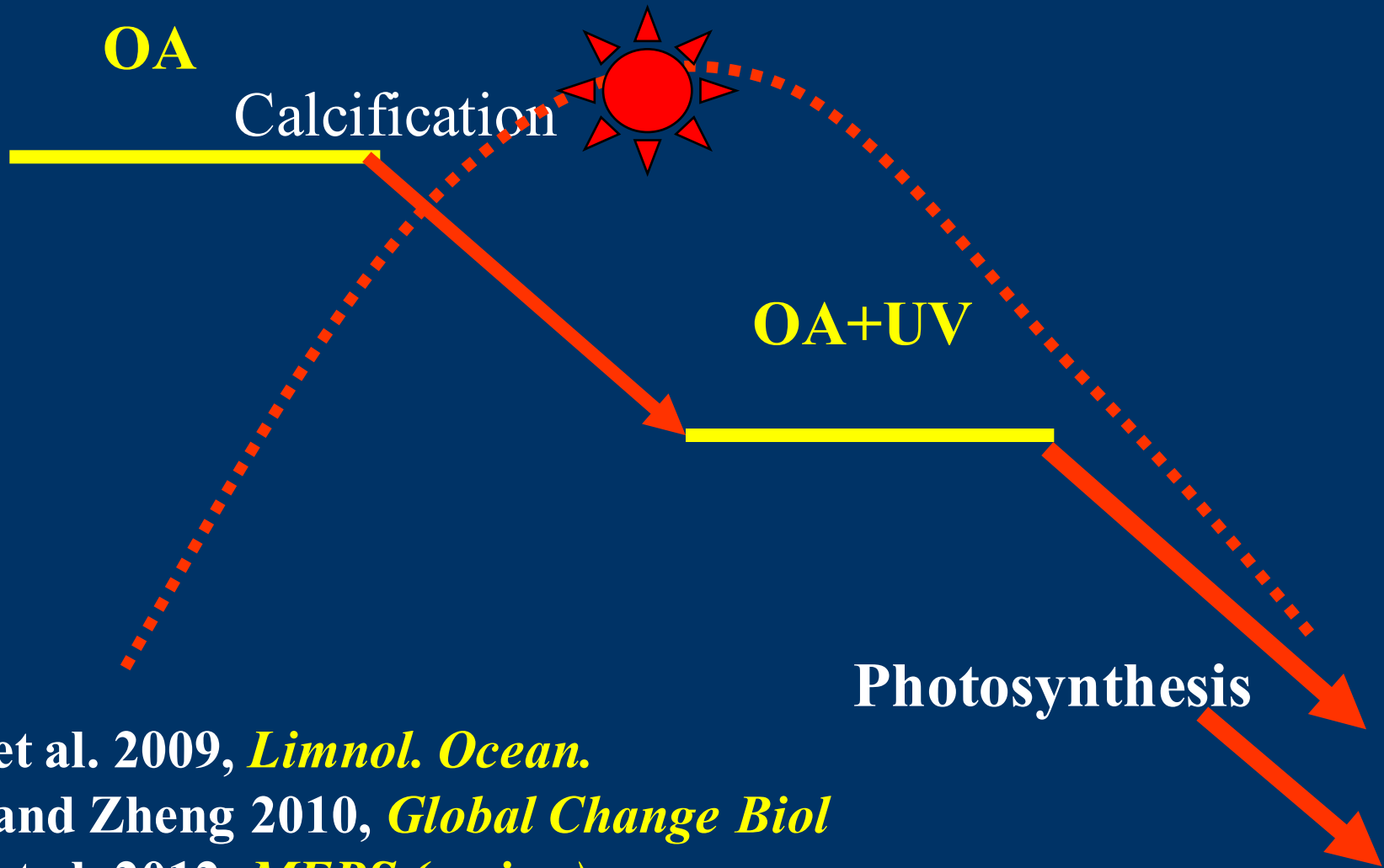


# Coraline algae



*E. hux*

# Synergistic impacts of OA and UV on calcifying algae



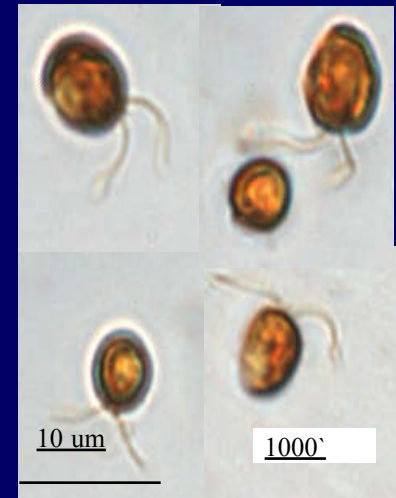
Gao et al. 2009, *Limnol. Ocean.*

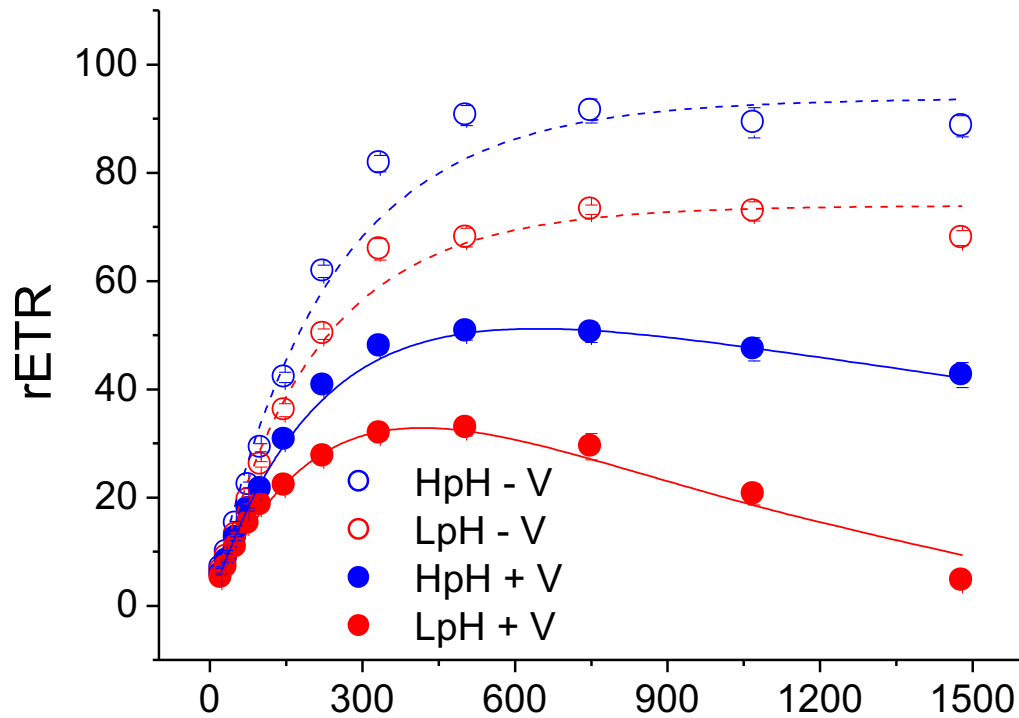
Gao and Zheng 2010, *Global Change Biol*

Gao et al. 2012 *MEPS (review)*

## Virus as a bio-stressor

Ocean acidification influences the effects of viral infection on a bloom-forming alga





赤潮藻（棕囊藻）病毒  
恶化该藻受酸化的影响

Global Change Biology

**Viral attack exacerbates the susceptibility of a bloom-forming alga to ocean acidification**

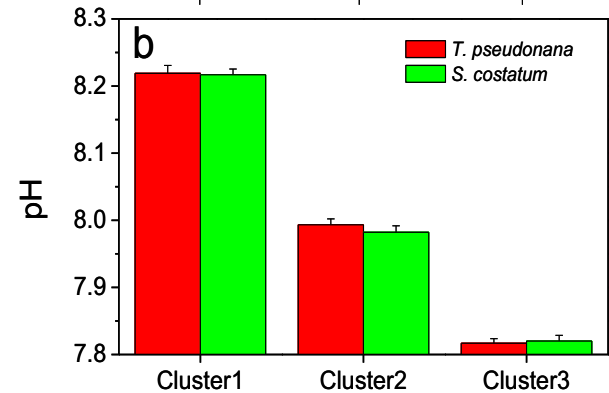
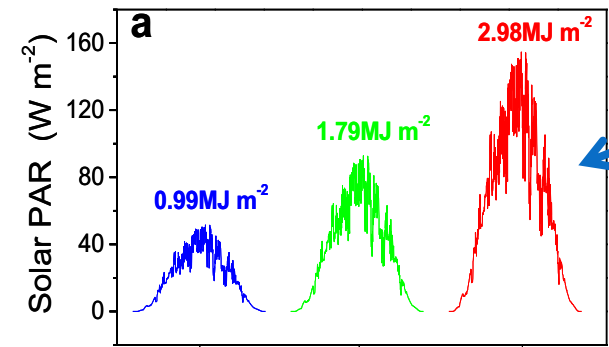
SHANWEN CHEN<sup>1</sup>, KUNSHAN GAO<sup>2</sup> and JOHN BEARDALL<sup>3</sup> 2014

# Enhanced Stratification

Less nutrients

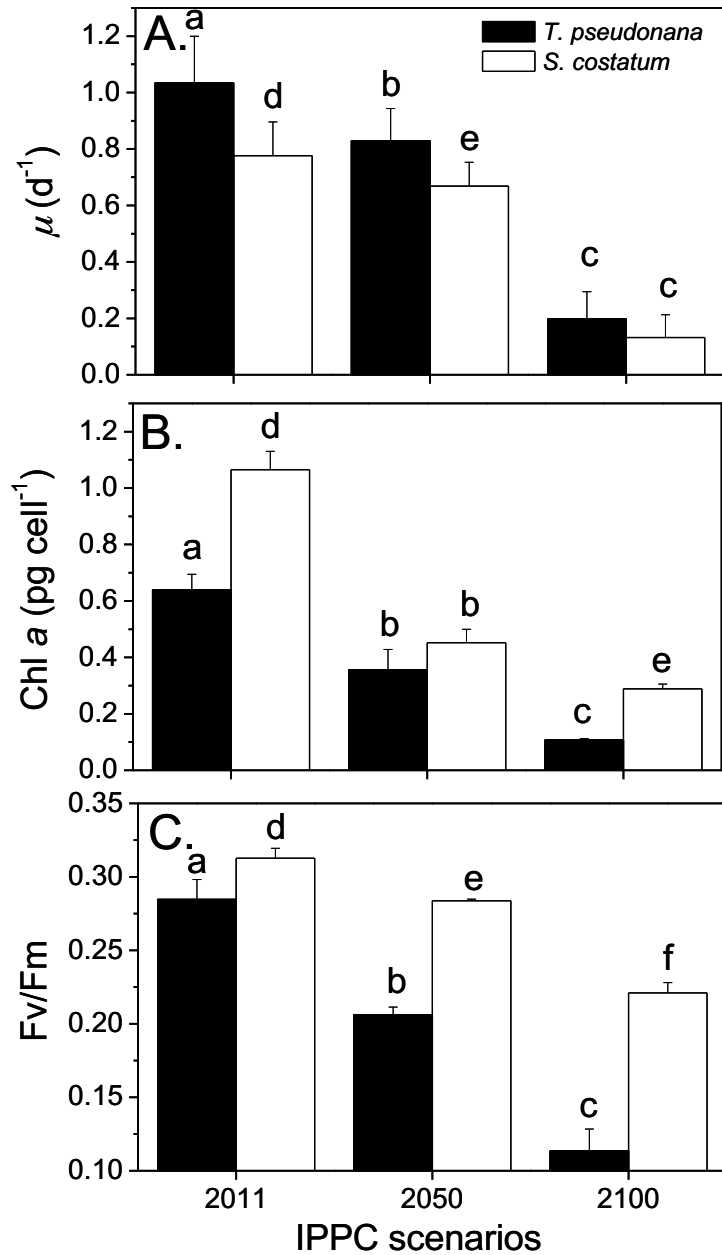
Increased light exposures

# Ocean acidification



**2011 2050 2100**

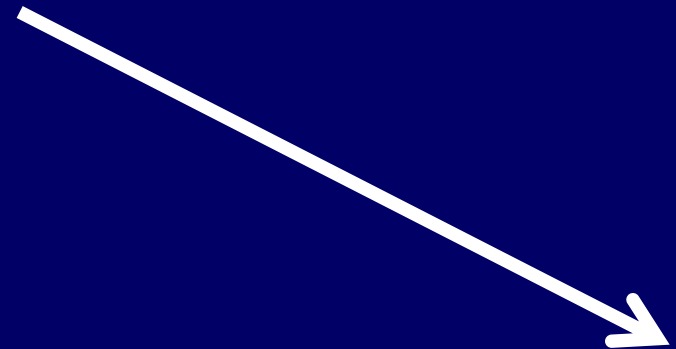
Scenarios	Light (% of surface)	CO <sub>2</sub> ( $\mu\text{atm}$ )	pH	NO <sub>3</sub> <sup>-</sup> ( $\mu\text{M}$ )	PO <sub>4</sub> <sup>3-</sup> ( $\mu\text{M}$ )	Si ( $\mu\text{M}$ )
2011	10%	390	8.18	20	2	20
2050	18%	700	7.96	10	1	10
2100	30%	1000	7.82	5	0.5	5



Time



Growth, Chl a, Fv/Fm



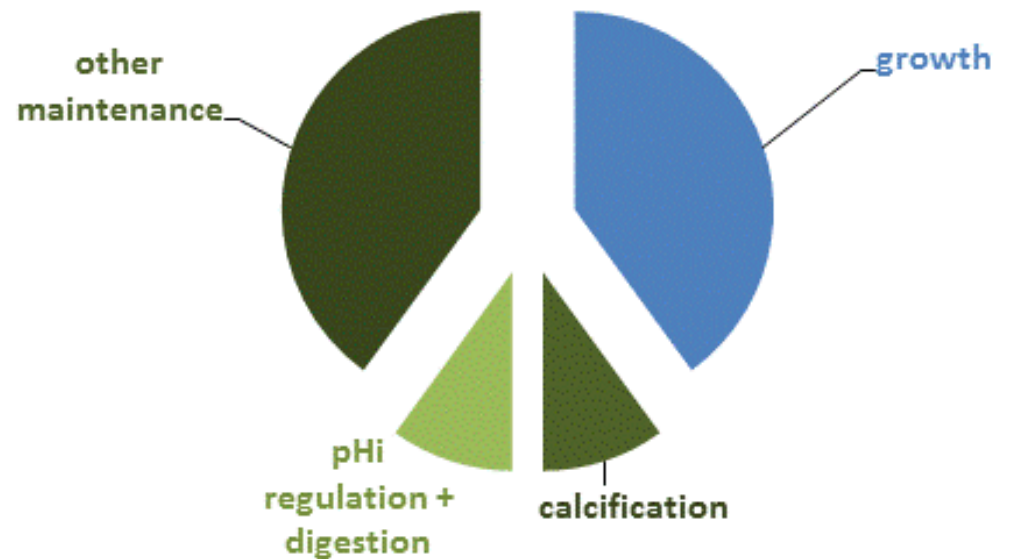
*Xu et al. 2014 MEPS*

# Shift in energy budget

control



ocean acidification



# Summary

- **+/- effects of OA depends on light or solar radiation levels**
- **Ocean acidification increases levels of phenolic compounds in phytoplankton and zooplankton, implying a food chain impact.**
- **UV radiation and OA interact synergistically to reduce algal calcification and photosynthetic C fixation**
- **Virus infection to primary producers would be worse under OA**





OA and taste of shrimps  
*Dupont et al. 2014*

**Thank you for your attention**

**Funded by**

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MOST  
SOA**

