



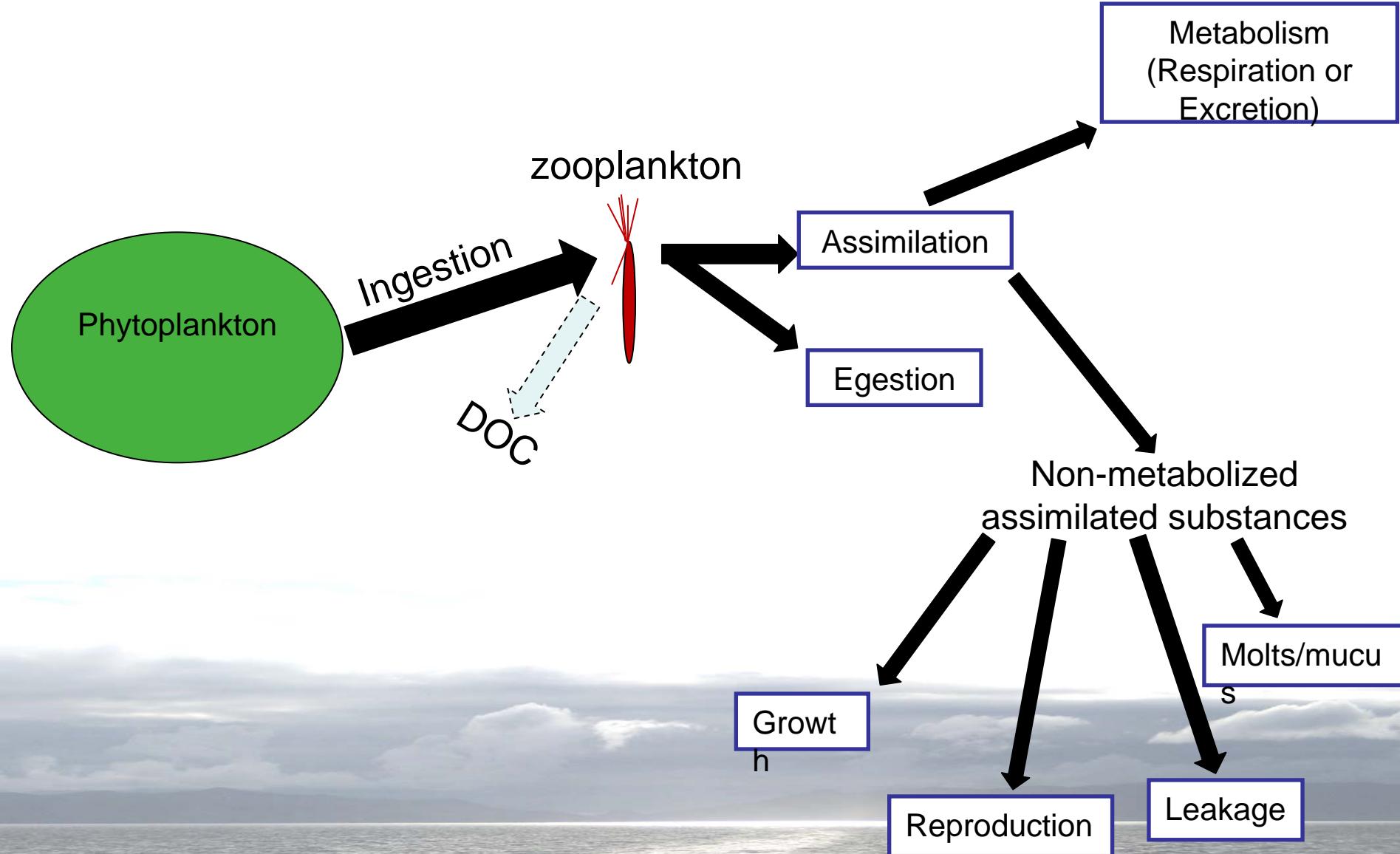
Microzooplankton feeding and growth in an acidified ocean

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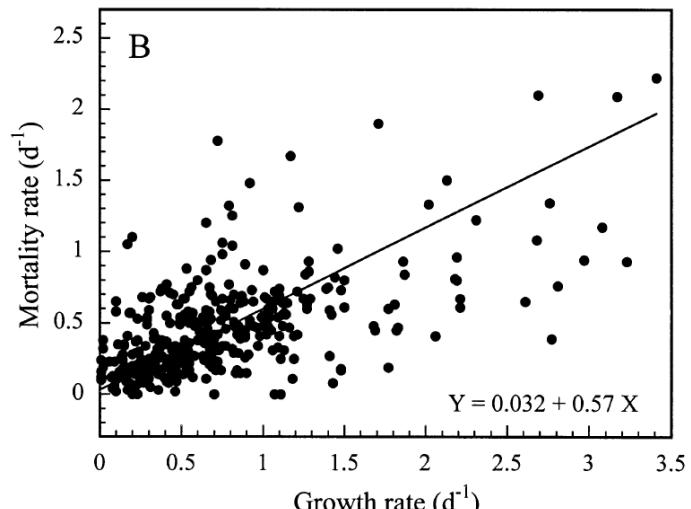
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- Ingestion

- Prey abundance
 - e.g. Verity, P. G. 1985; Jakobsen, H. H. & P. J. Hansen, 1997
- Prey growth rate
 - e.g. Calbet, A., Landry, M.R. 2004
- Prey nutritional quality (C:N:P)
 - e.g. Butler, N. M., C. A. Suttle & W. E. Neill, 1989; Cowles, T. J., R. J. Olson & S.W. Chishom, 1988
- Prey size
 - e.g. Gonzalez, J. M., E. B. Sherr & B. F. Sherr, 1990; Jakobsen, H. H. & P. J. Hansen, 1997
- Prey defense mechanisms
 - e.g. Strom, S.L., G.V. Wolfe, A. Slajer, S. Lambert, and J. Clough. 2003

- Assimilation (egestion)
 - Prey abundance (high or low)
 - e.g. Landry et al. 1984
 - Prey nutritional quality (C:N:P)
 - e.g. Mitra and Flynn 2005



From Calbet, A., Landry, M.R. 2004

Microzooplankton will be indirectly affected by ocean acidification (elevated $p\text{CO}_2$) through...

I. Alterations in physiology, rate processes and biochemistry of phytoplankton

- i. Photosynthetic rate
- ii. C:N:P stoichiometry
- iii. Size, cell division rate, defense properties
- iv. DOC/TEP release

II. Alterations in zooplankton ecology

- i. Feeding and growth rates
- ii. Excretion rate, products, stoichiometry

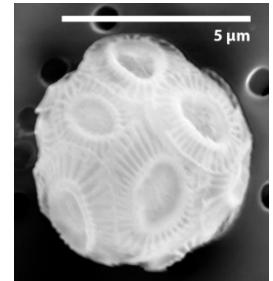
Possible consequences:

Change in food web efficiency

Shift to microbially dominated food web

Design and Methods:

- Grow phytoplankton cultures semi-continuously in lab across a range of $[p\text{CO}_2]$
 - Model phytoplankton: calcifying (CCMP 2668) and non-calcifying (CCMP 374) *Emiliania huxleyi*
 - $[p\text{CO}_2]$ of ambient (~395), 750 and 1000 $p\text{CO}_2$
 - Measure: cell growth rate, C:N, chlorophyll a, photosynthetic rate, DMSP, PIC:POC, cell size, calcification rate, coccolith morphology
- Feed acclimated *E. huxleyi* to microzooplankton
 - Model microzooplankton: *Amphidinium longum*, *Oxyrrhis marina*, *Eutintinnis* sp., *Strombidinopsis* sp., more to come...
 - Measure: grazing rate, ingestion rate, growth rate



Shear affects ‘normal’ microzooplankton behavior:

- Mortality

Laboratory culture of marine planktonic oligotrichs (Ciliophora, Oligotrichida)

Dian J. Gifford 1985

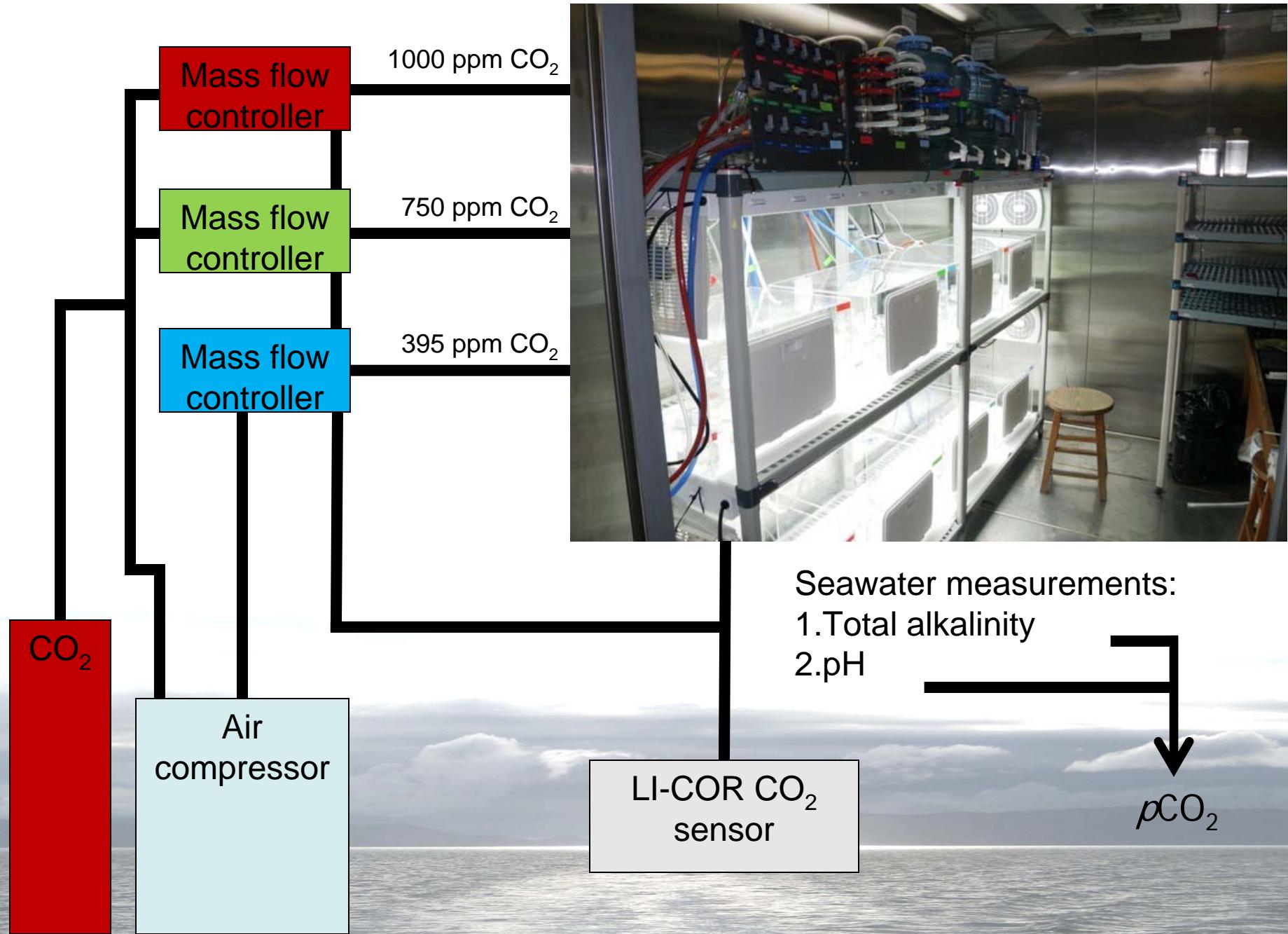
Conserving original *in situ* diversity in microzooplankton grazing set-ups

Martin Günter Joachim Löder et al. 2010

- Feeding behavior

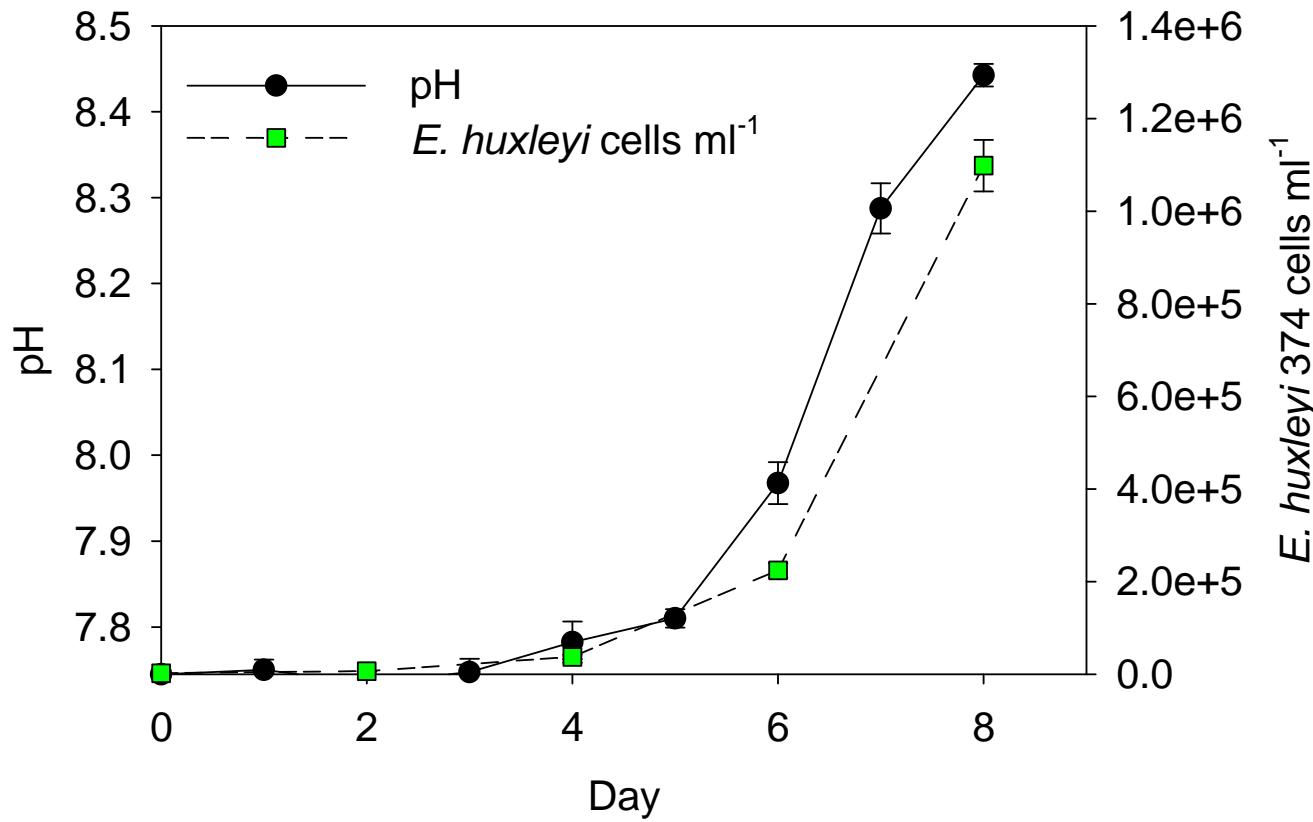
Influence of turbulence on suspension feeding by planktonic Protozoa; experiments in laminar shear fields

Martin Günter Joachim Löder et al. 2010



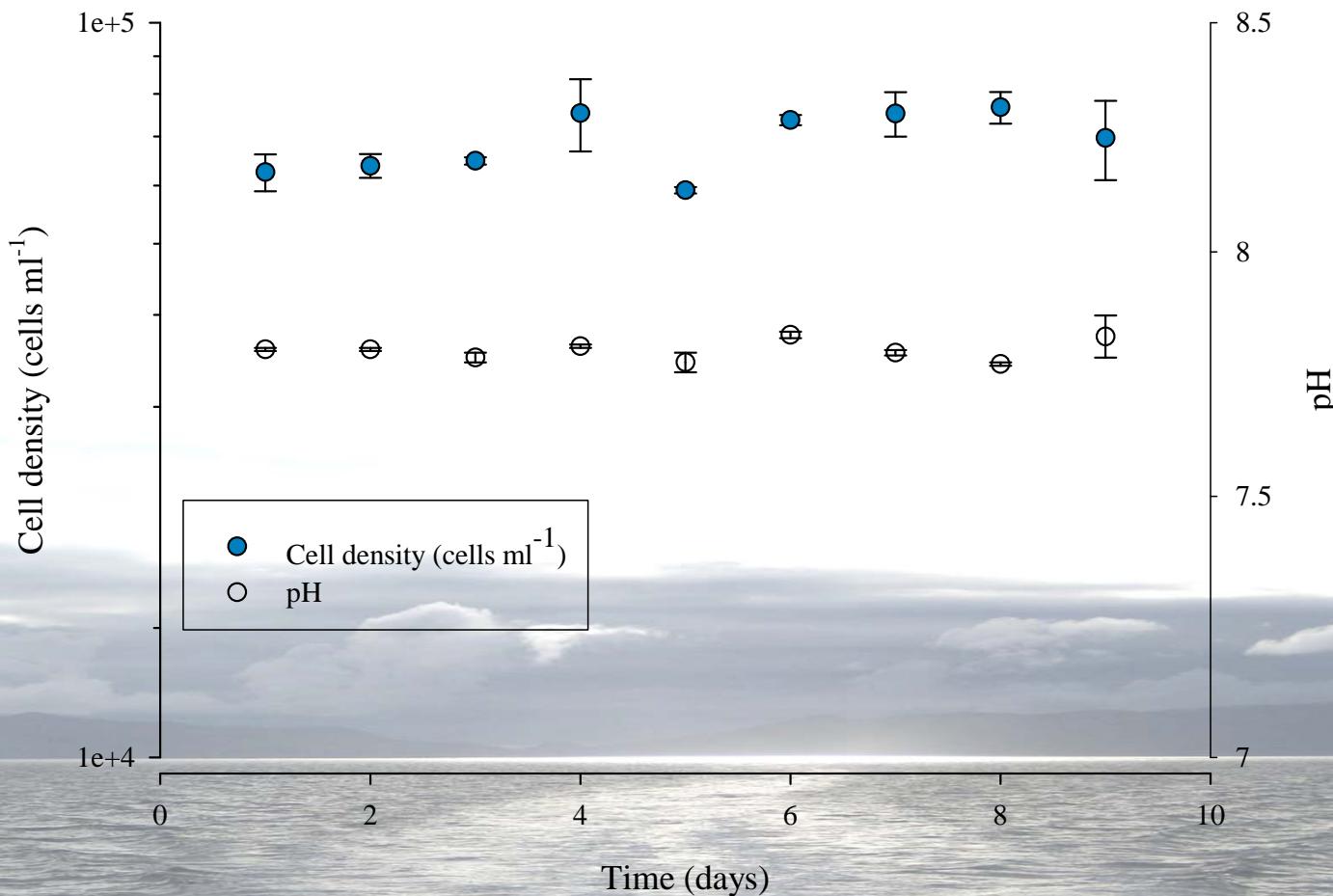
E. Huxleyi biomass eventually consumes CO₂ faster than it is supplied through gas exchange

- Conditions: *E. huxleyi* (CCMP 374) grown at 1000 pCO₂
- Daily cell counts (FC) and pH measurements (Metrohm 888 Titrando)



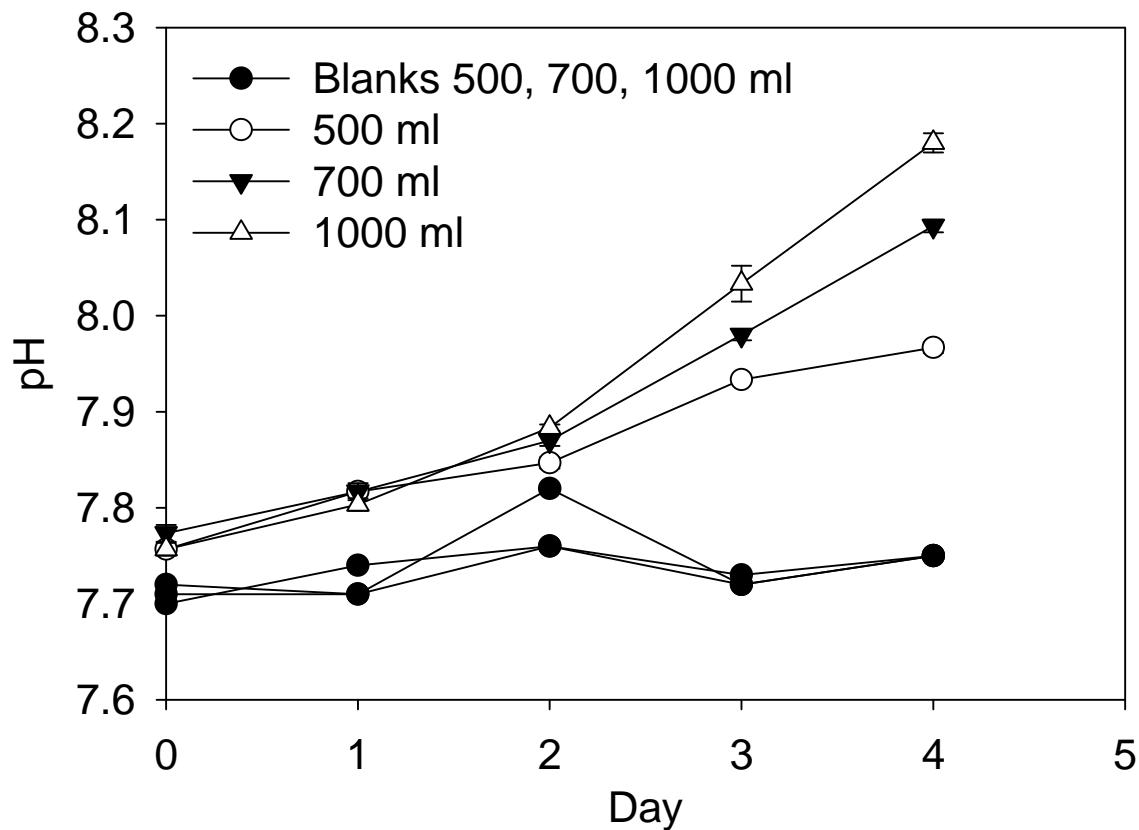
At cell concentrations $\leq 100,000$ cells ml^{-1} CO_2 chemistry maintained

- Conditions: *E. huxleyi* (CCMP 374) grown at 1000 pCO_2
- Daily cell counts (FC) and pH measurements (Metrohm 888 Titrando)



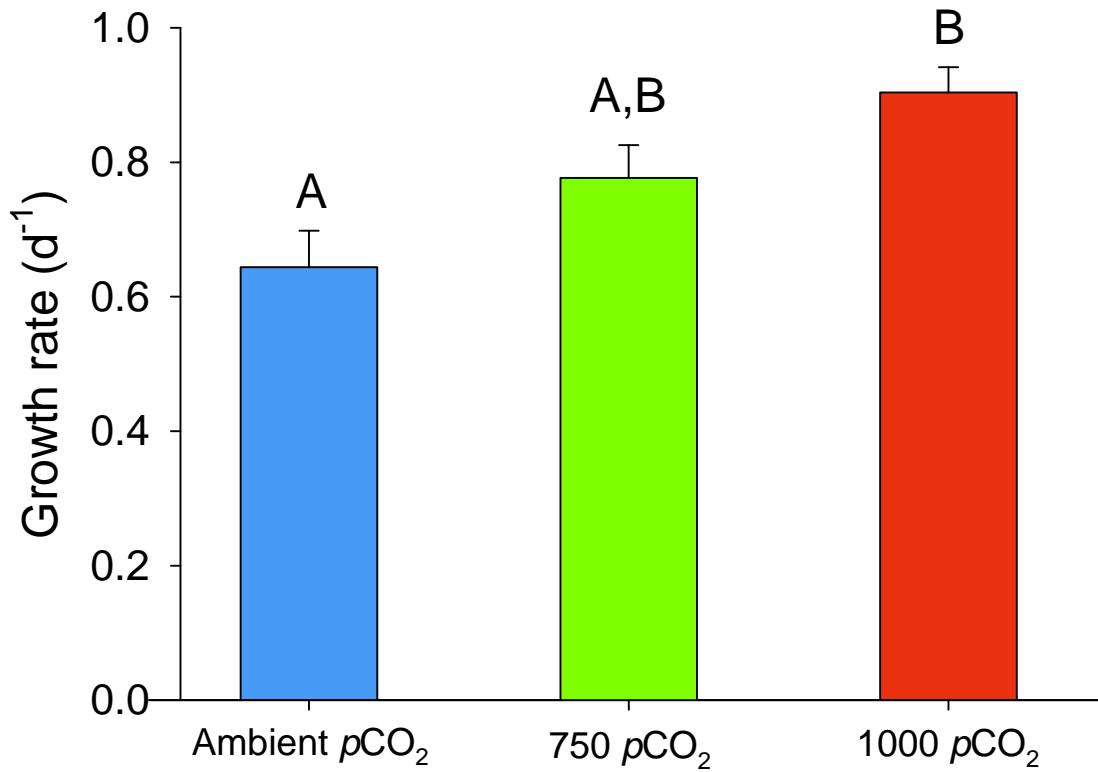
Can we find surface area:volume ratio that balances sample volume needs with gas exchange?

- Conditions: *E. huxleyi* (CCMP 2668) grown at 1000 $p\text{CO}_2$
- Daily cell counts (FC) and pH measurements (Metrohm 888 Titrando)



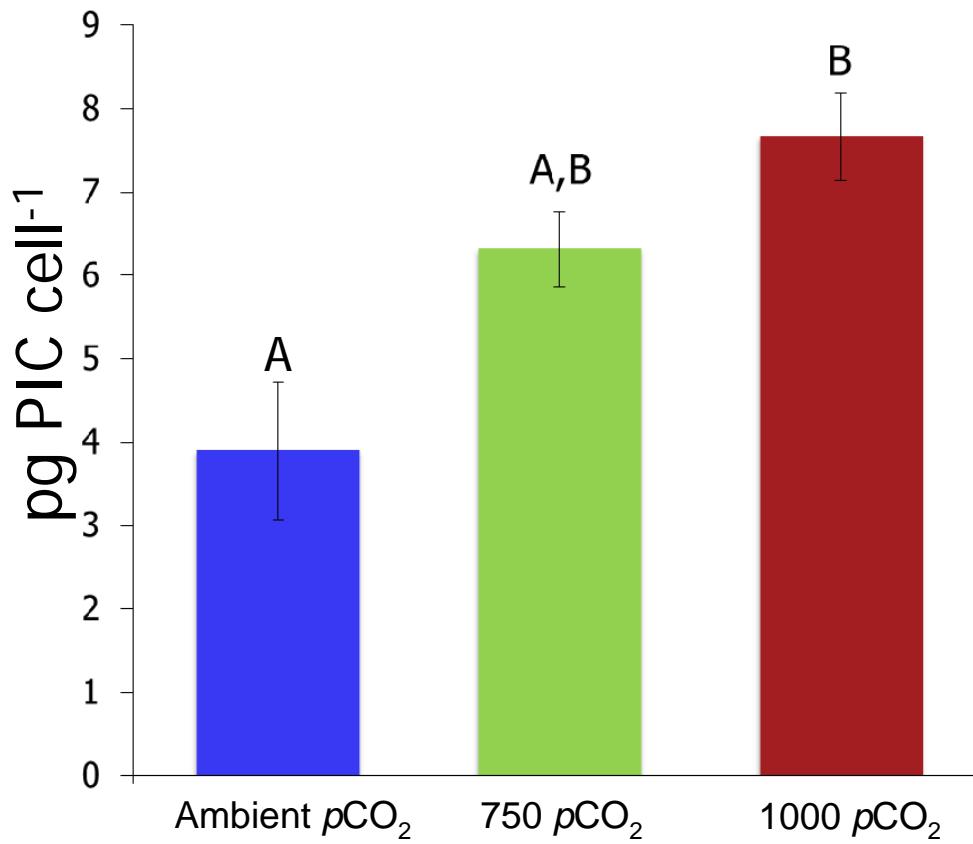
Physiological/biochemical responses of *E. huxleyi* (CCMP 2668) to elevated $p\text{CO}_2$

- Conditions: *E. huxleyi* (CCMP 2668) grown semi-continuously for 10 days at ambient, 750 and 1000 $p\text{CO}_2$
- Daily cell counts

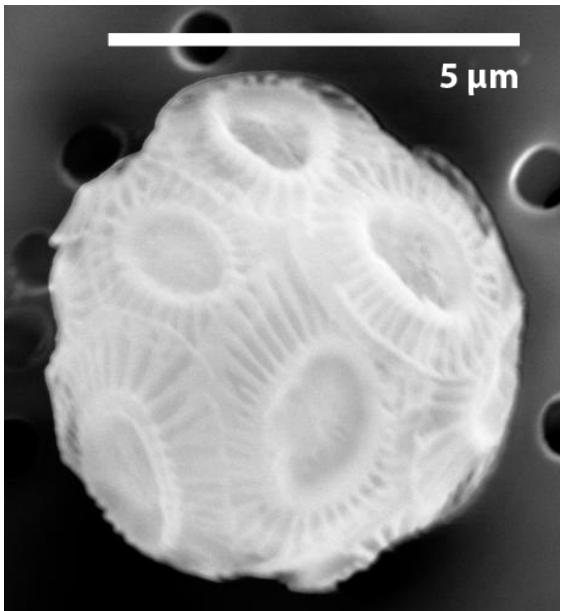


More PIC per cell at higher $p\text{CO}_2$

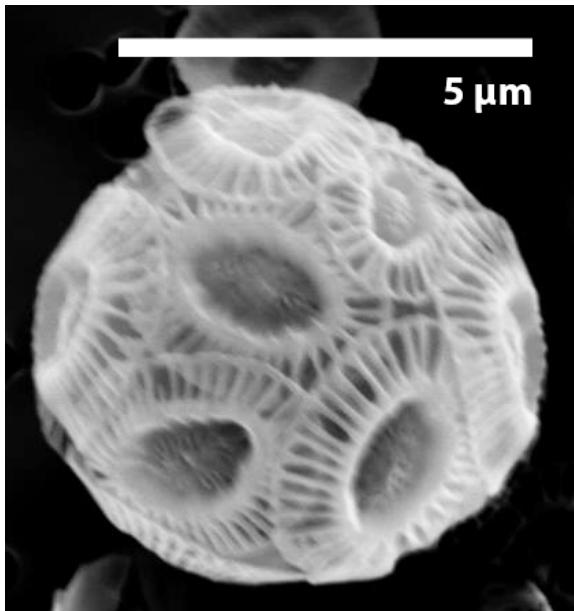
- Conditions: *E. huxleyi* (CCMP 2668) grown semi-continuously for 10 days at ambient, 750 and 1000 $p\text{CO}_2$
- Daily cell counts and PIC measurements day 8



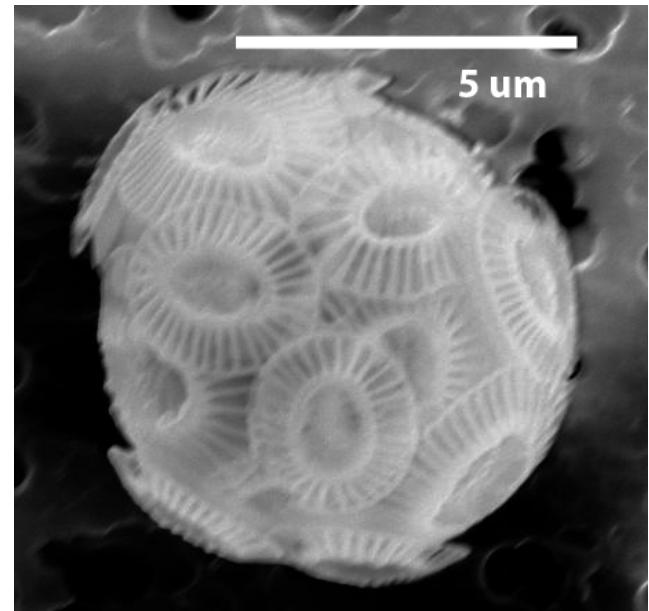
Increasing coccoliths cell^{-1} , cell diameter (2668)



Ambient
(395 pCO_2)



Moderate
(750 pCO_2)



High
(1000 pCO_2)

Experimental conditions:

- *Oxyrrhis marina* acclimated (8 days) to 3 [pCO₂]
- *O. marina* fed non-acclimated *E. huxleyi*
- *O. marina* short-term ingestion and growth (24h) measured

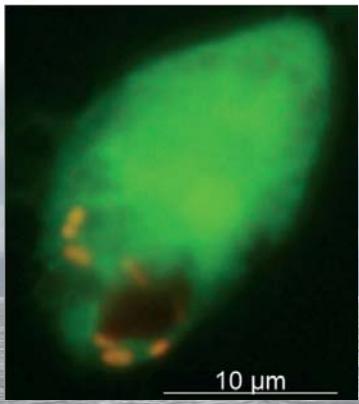
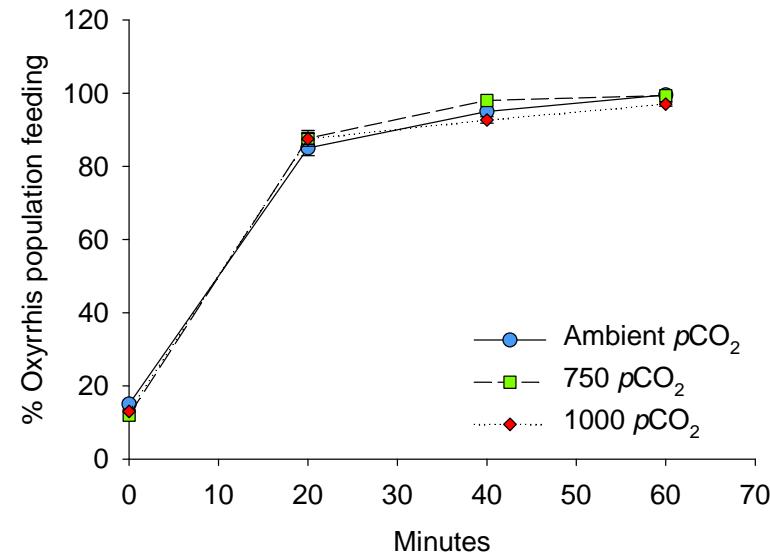
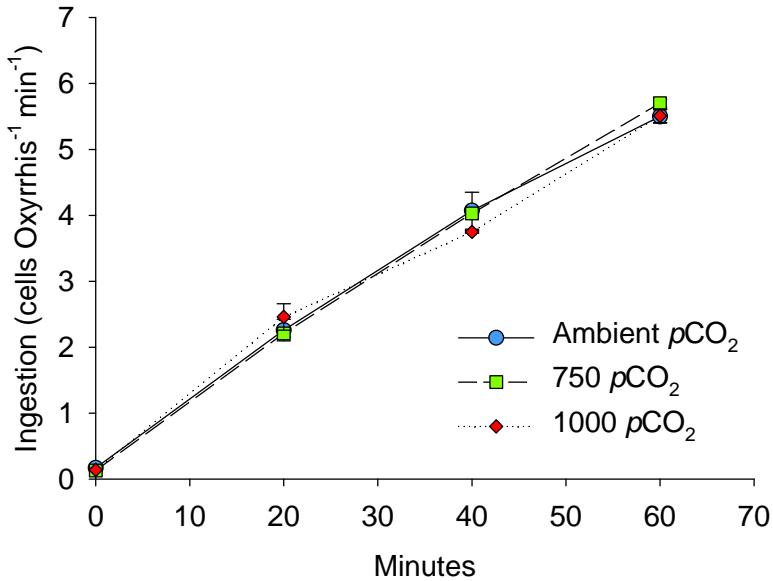
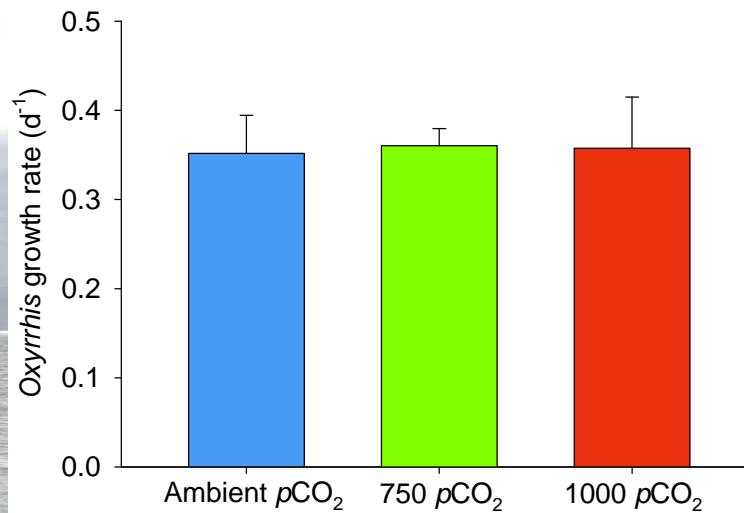
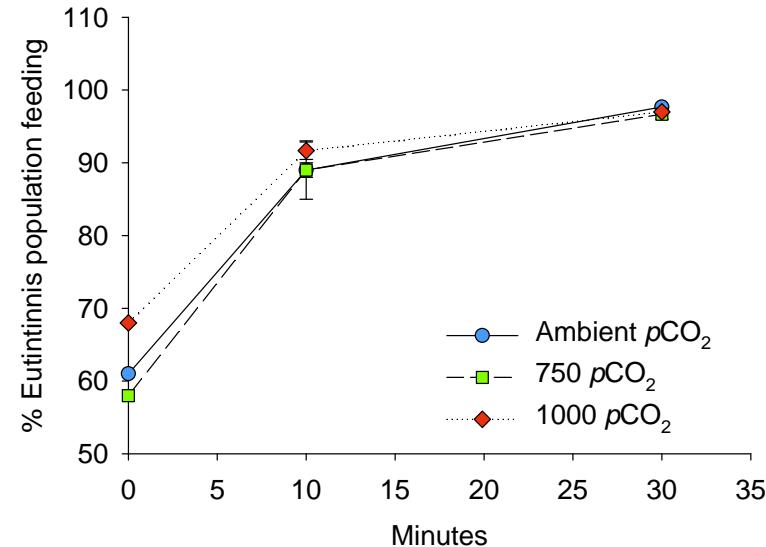
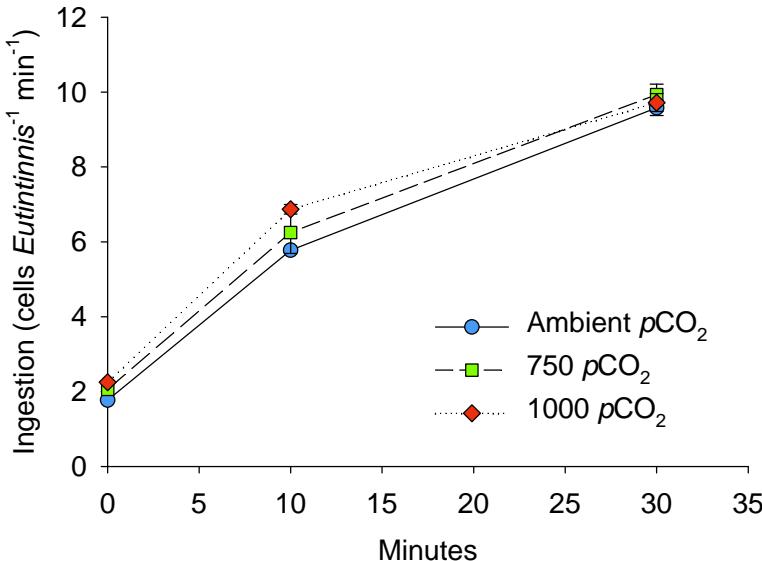


Photo: Jude Apple

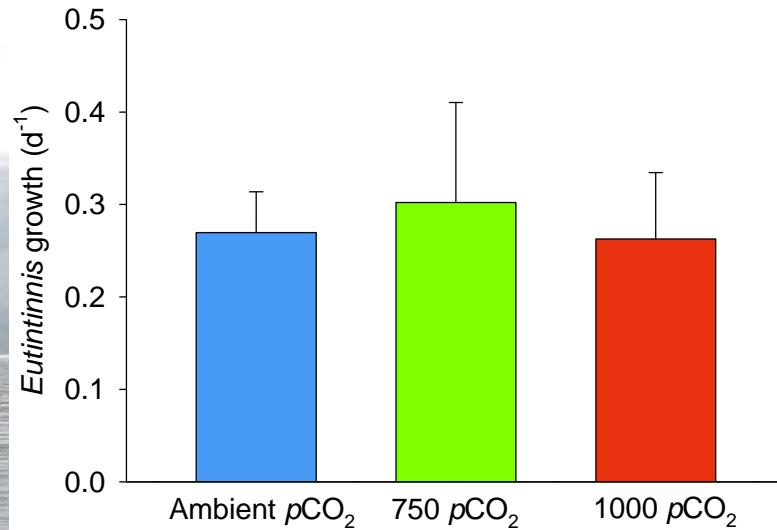


Experimental conditions:

- *Eutintinnus* sp. acclimated (8 days) to 3 [pCO₂]
- *Eutintinnus* sp. fed non-acclimated *E. huxleyi* CCMP 374
- *Eutintinnus* sp. short-term ingestion and growth (24h) measured



<http://forum.mikroskopia.com/index.php?showtopic=4907>



Experimental conditions:

- *Amphidinium longum* acclimated (8 days) to 3 [pCO₂]
- *A. longum* fed non-acclimated *E. huxleyi* CCMP 374
- *A. longum* short-term ingestion and growth (24h) measured

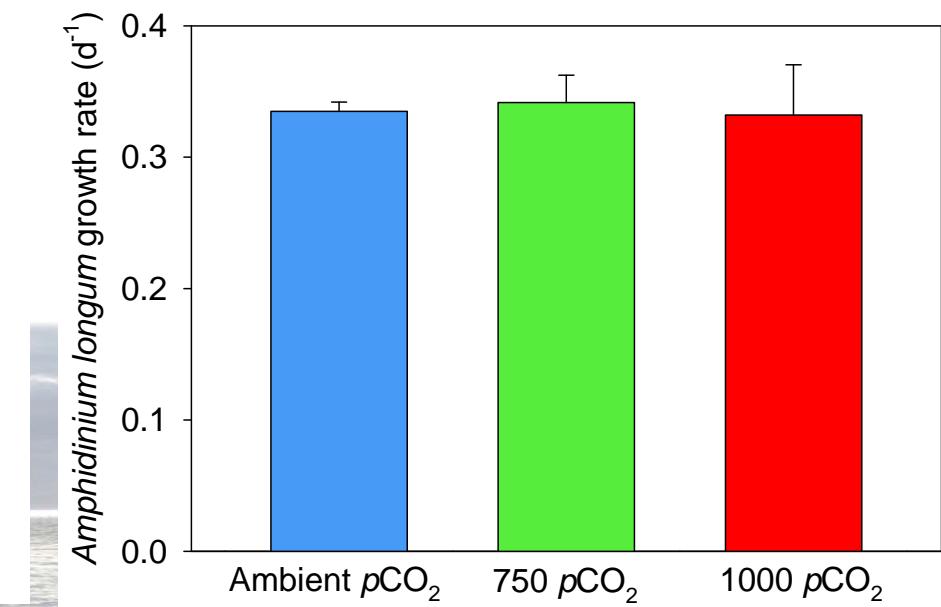
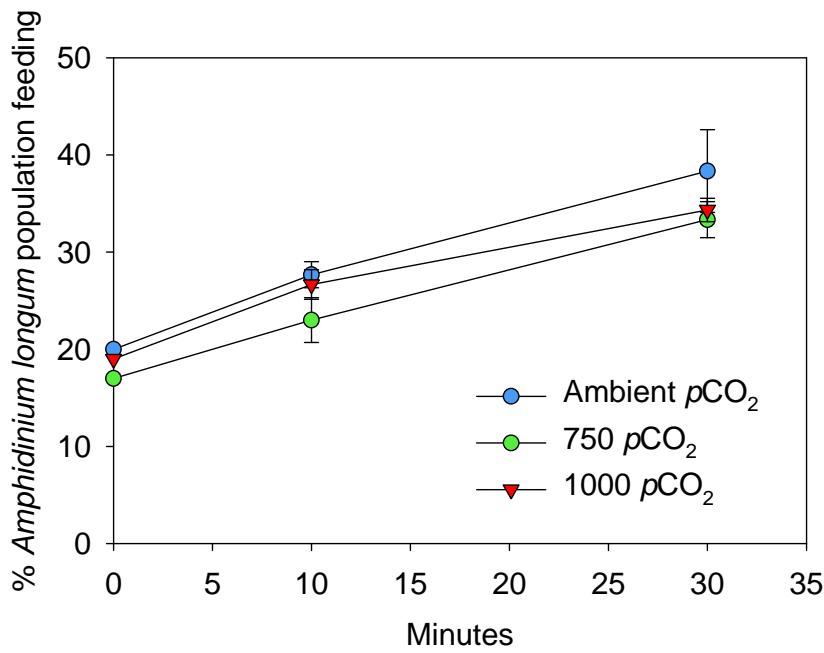
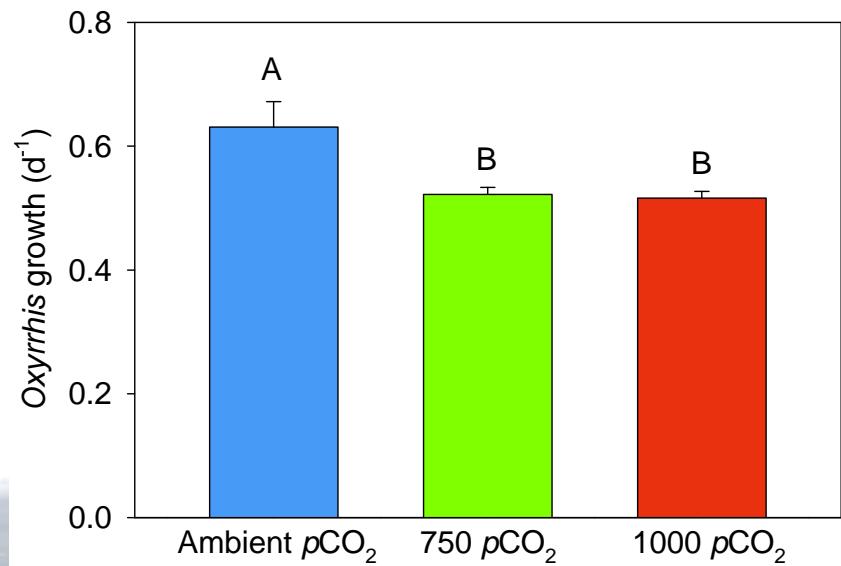
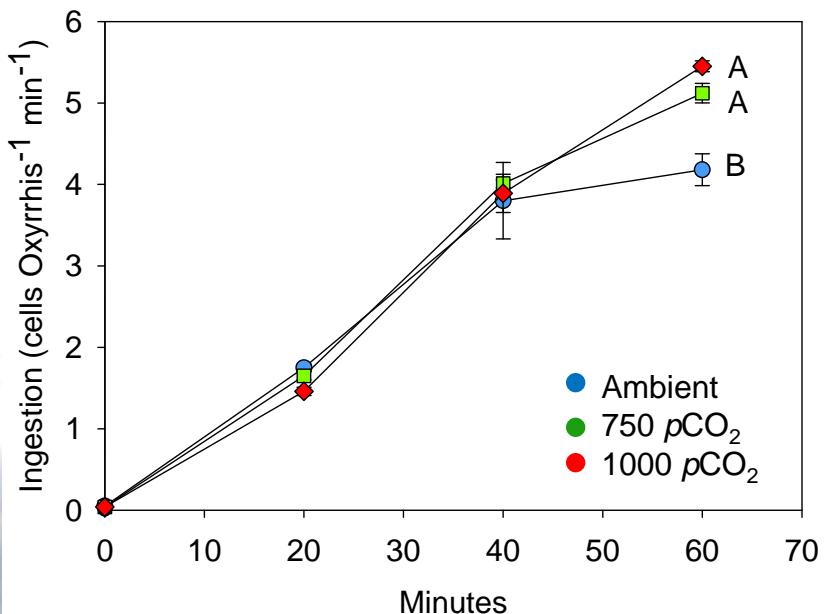


Photo: Gordon V. Wolfe

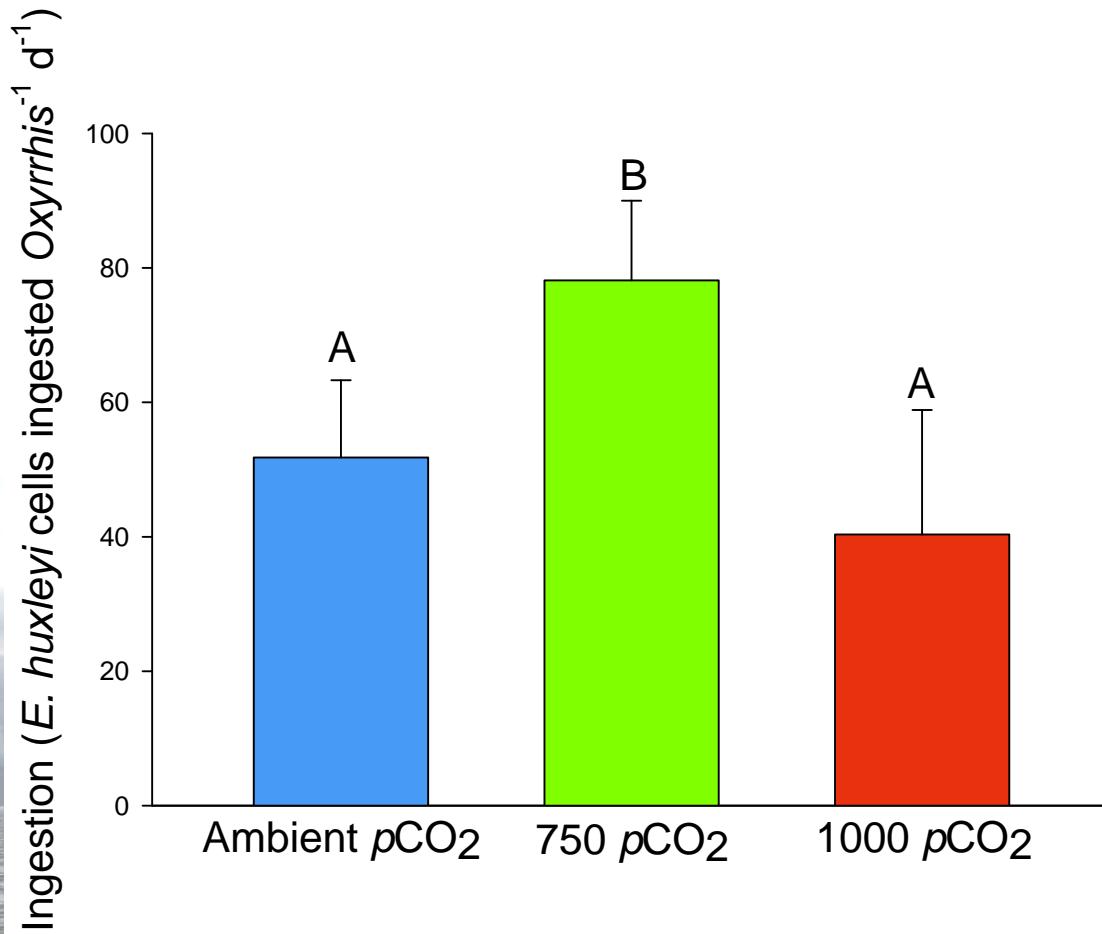
Experimental conditions: Testing for indirect effects

- *E. Huxleyi* 374 – grown semi-continuously for 8 days at 3 [pCO₂]
- pCO₂ acclimated *E. Huxleyi* 374 fed to non-acclimated *O. marina*
- *O. marina* short-term ingestion and growth (48h) measured



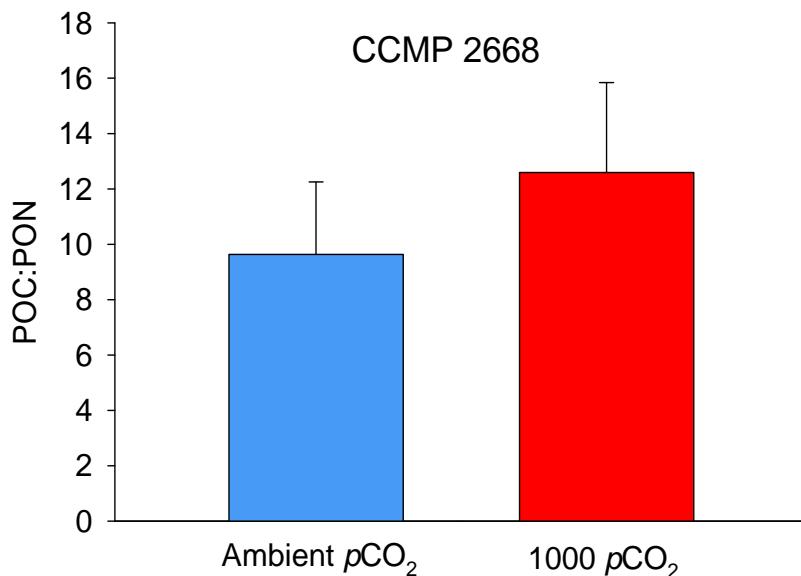
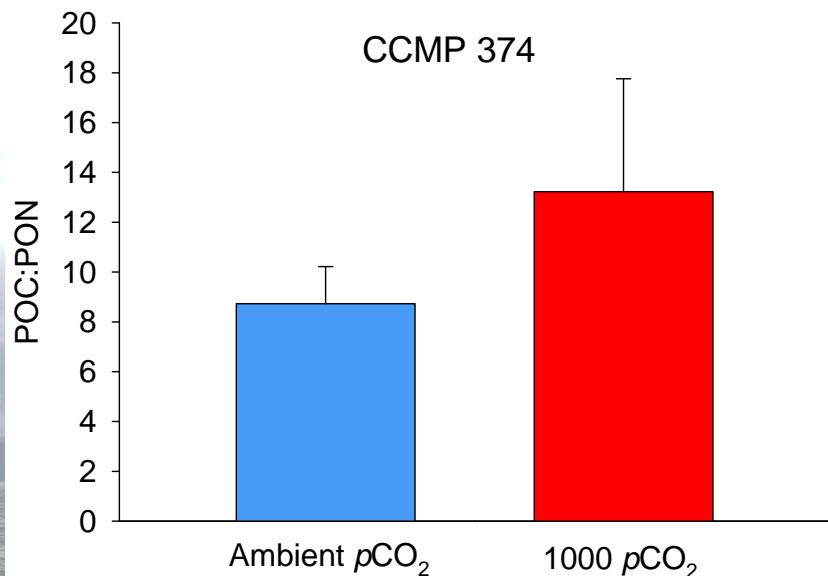
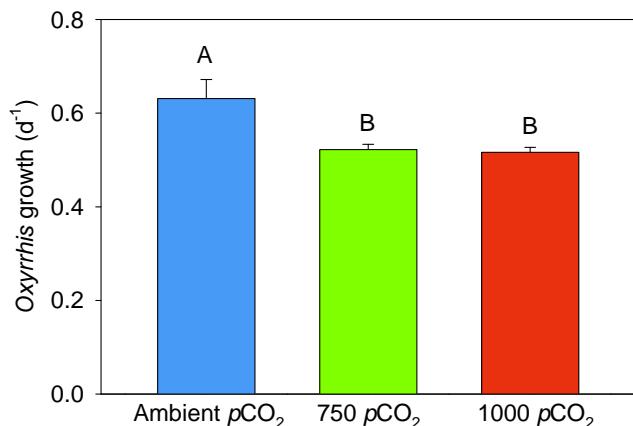
Experimental conditions:

- *E. Huxleyi* 374 – grown semi-continuously for 8 days at 3 [pCO₂]
- pCO₂ acclimated *E. Huxleyi* 374 fed to non-acclimated *O. marina*
- *O. marina* ingestion measured by cell disappearance (24h)



Experimental conditions:

- *E. Huxleyi* CCMP 374 and 2668 grown semi-continuously for 10 days at 3 [pCO₂]
- C:N measured on days 1, 6 and 10
- Averaged C:N for days 1, 6 and 10



Conclusions:

- There is no conspicuous direct effect of elevated $p\text{CO}_2$ to microzooplankton
- This study and others show physiological and biochemical responses by phytoplankton to elevated $p\text{CO}_2$
- Microzooplankton may alter their feeding behavior in response to $p\text{CO}_2$ -induced changes in phytoplankton
- Changes in microzooplankton feeding ecology will affect many important ocean processes

Thanks to students, SPMC, WWU, and NSF OCE 0961229